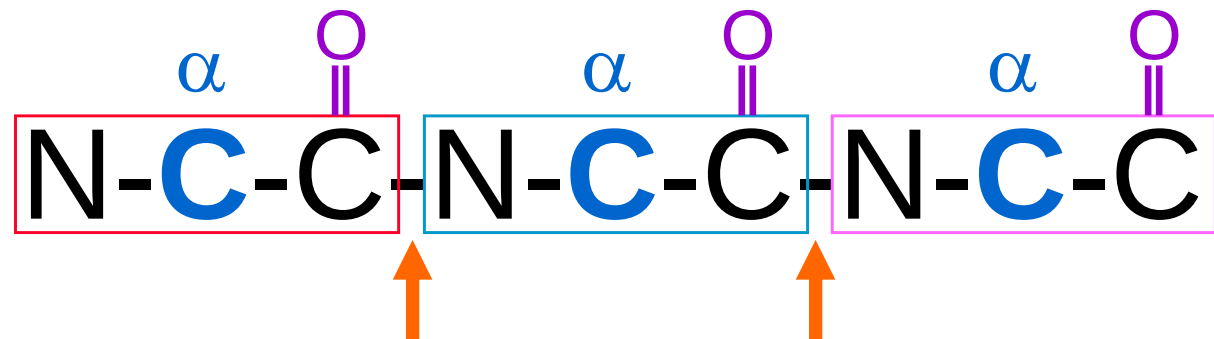
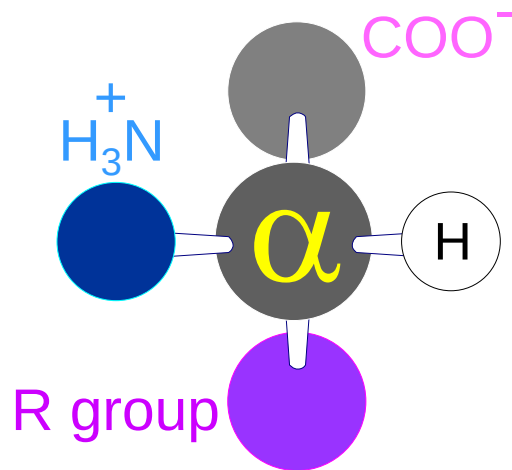
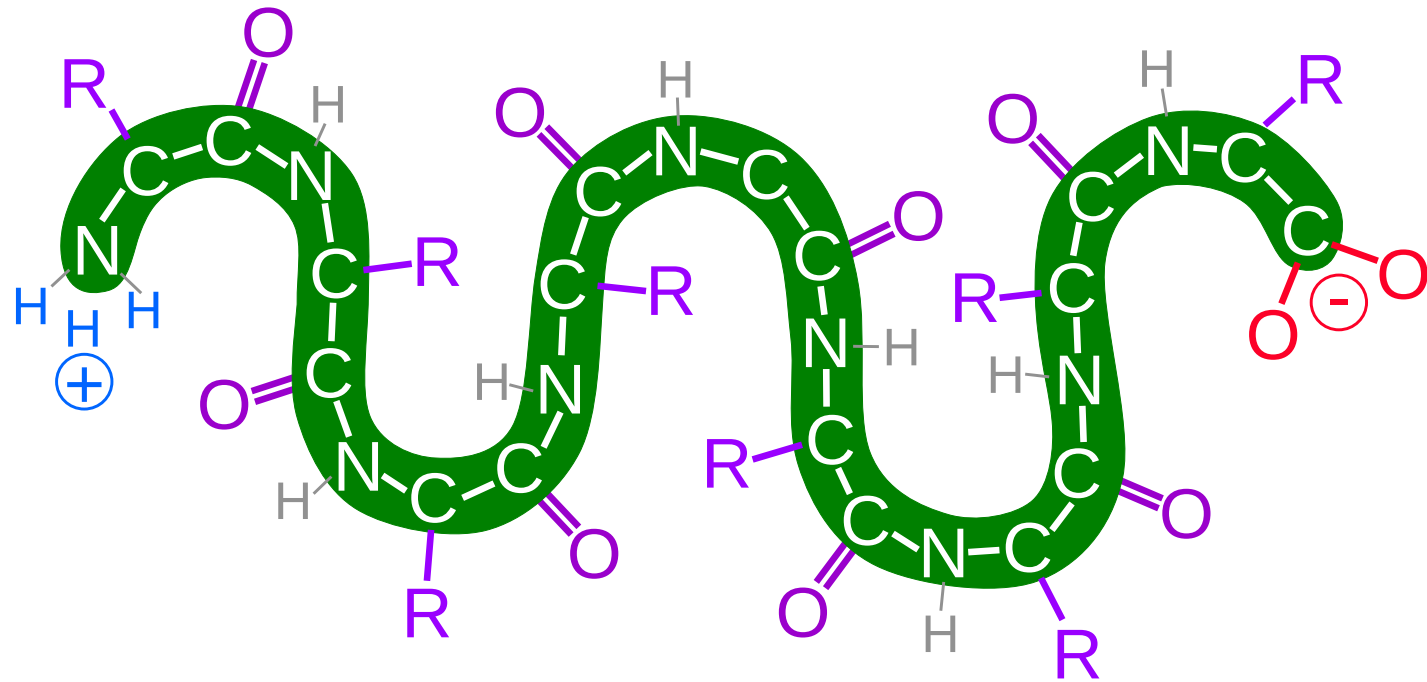


酵素分析方法



- 蛋白質定量 也是根基於分子構造
- 酵素活性分析 有催化活性的巨分子
- 膠體電泳法 SDS vs Native-PAGE
- 免疫轉印法 應用抗體的專一性
- Proteomics 解開基因序列之後

蛋白質構造的骨架



各種蛋白質定量法原理

5 Specific Binding Group

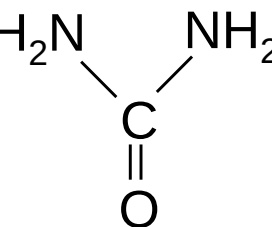
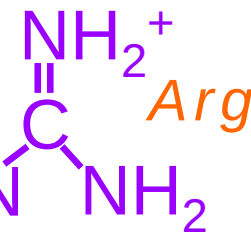
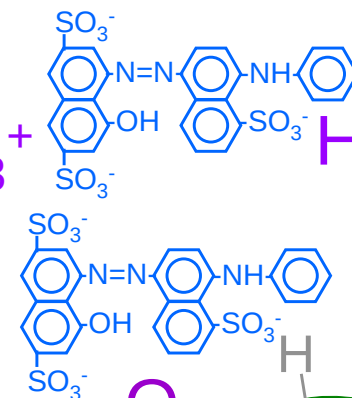
Heme



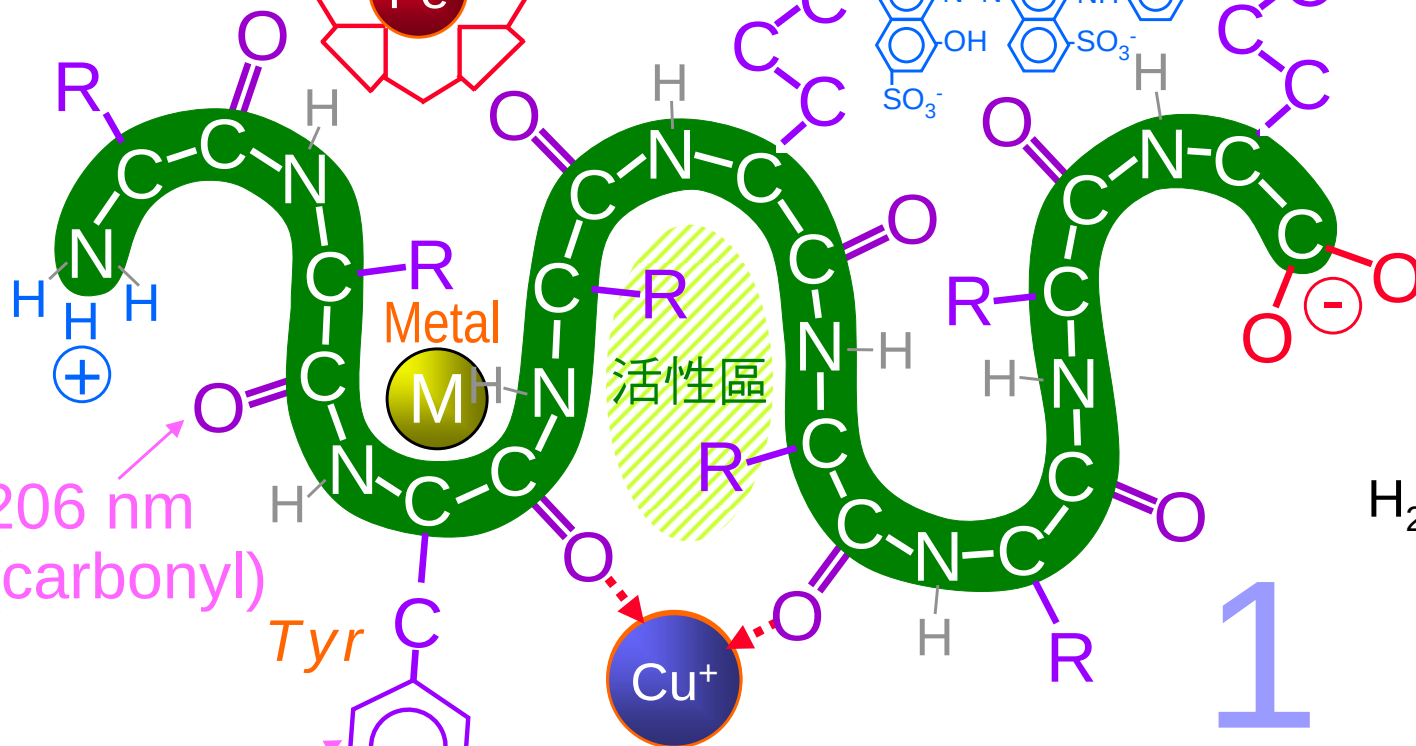
4

Coomassie Brilliant Blue G

Lys



尿素
urea



206 nm
(carbonyl)

Tyr

3

UV

Absorbance

280 nm
(aromatic)



Phosphomolybdic-phosphotungstate

Biruet Methods
(carbonyl)

Lowry Methods

1

2

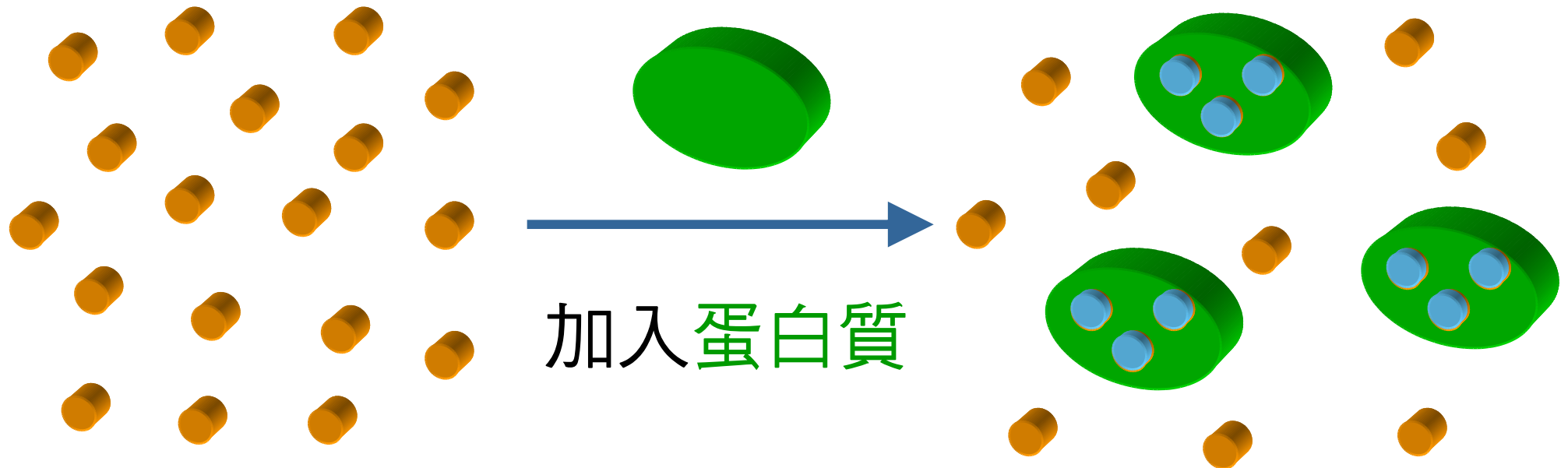
Bradford Method

Coomassie Brilliant Blue G-250

470 nm

CBG 是一種指示劑

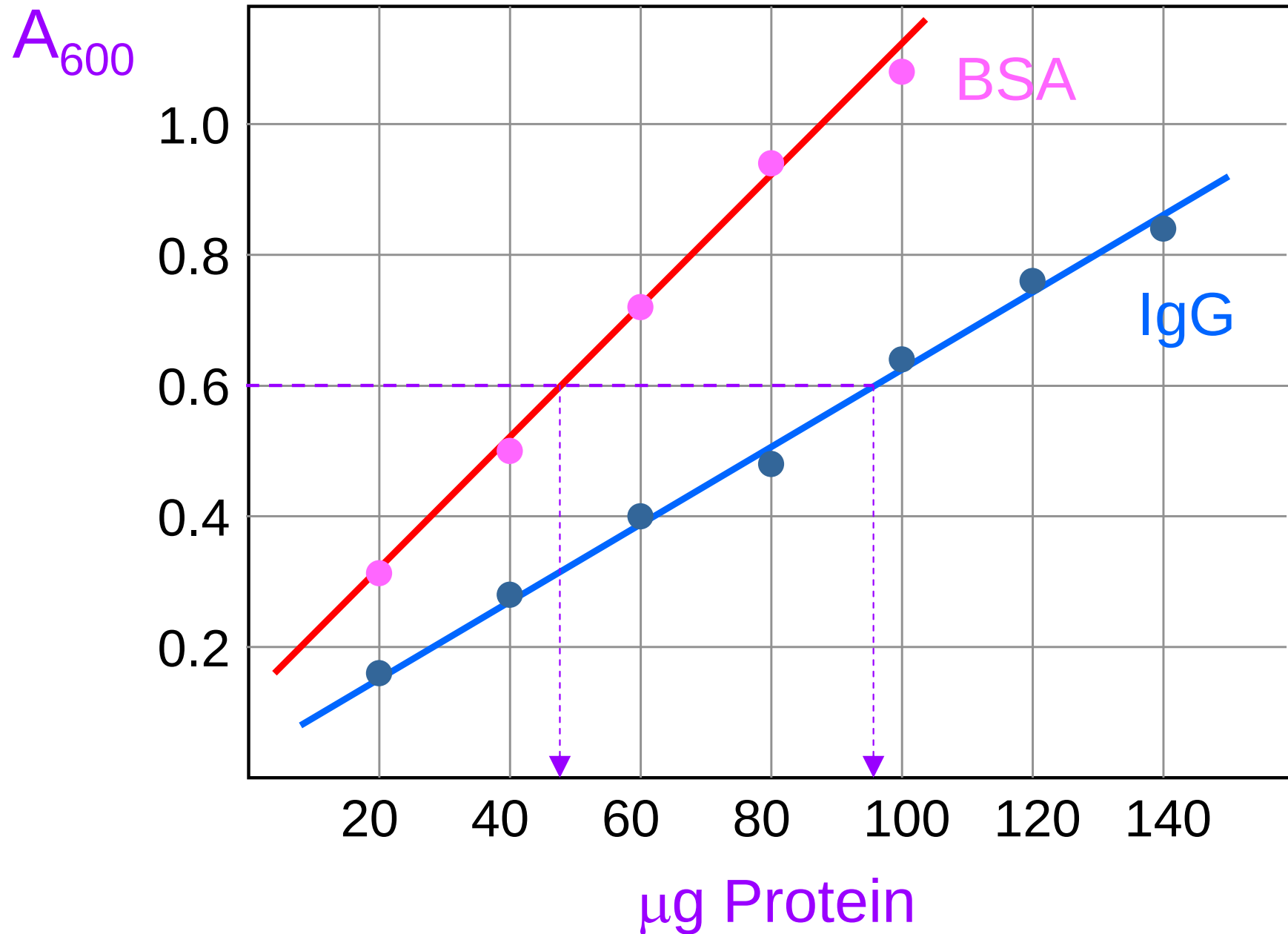
595 nm



酸性環境下呈茶色

與蛋白質結合變藍色

不同蛋白質的定量差異



■ 各種蛋白質定量法的比較

定量方法	靈敏度	準確度	其它說明
Biuret	0.05 - 5 mg	高	呈色快速 有腐蝕性 銨離子干擾大
Lowry	0.05 - 0.5 mg	中	呈色較慢 多種離子有干擾
Absorbance 280 nm	0.05 - 2 mg	低	樣本可回收 其它物質干擾大
Absorbance 205 nm	0.01 - 0.05 mg	高	樣本可回收 氧分子干擾極大
Bradford Dye-binding	0.01 - 0.05 mg	中 高	呈色快 雜質干擾多 顏色不易洗除

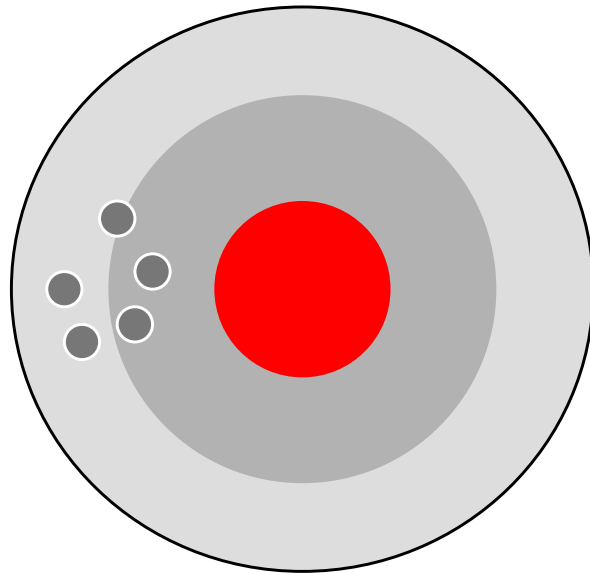
精確+準確

Bradford
Method

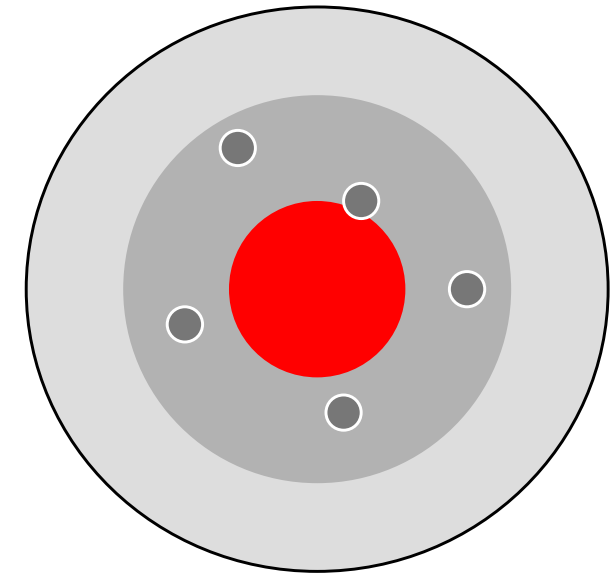
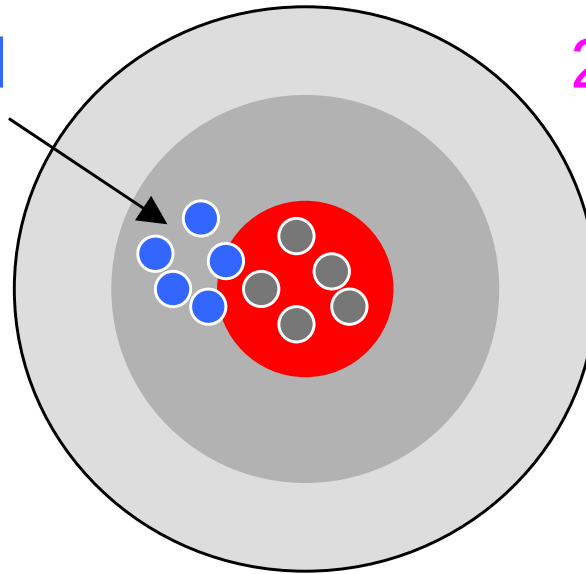
206 nm 吸光度

精確

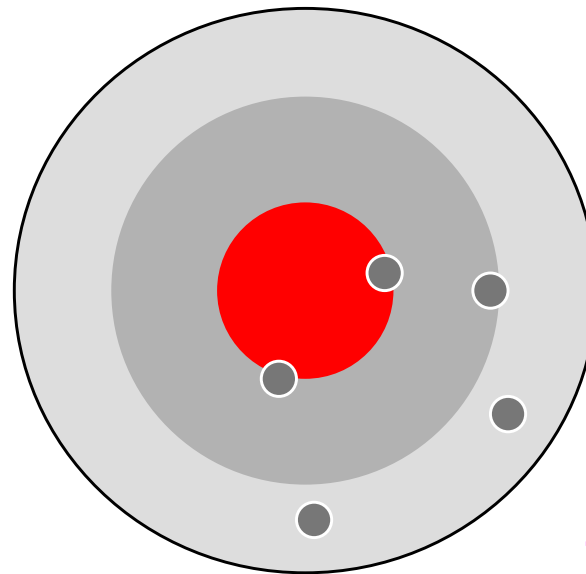
準確



Lowry Method



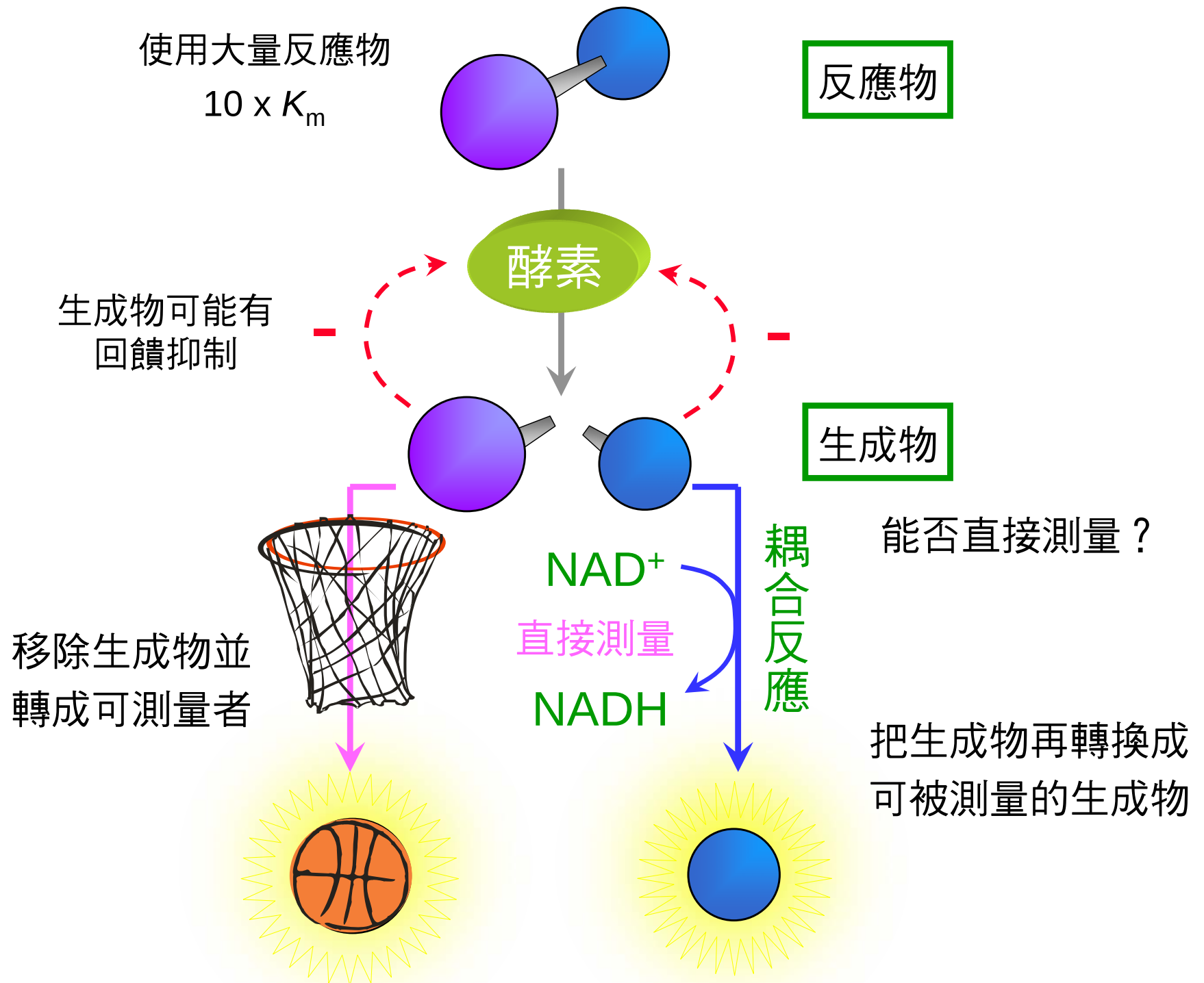
Biuret Method



280 nm 吸光度

不精確+不準確

酵素反應及偵測方法

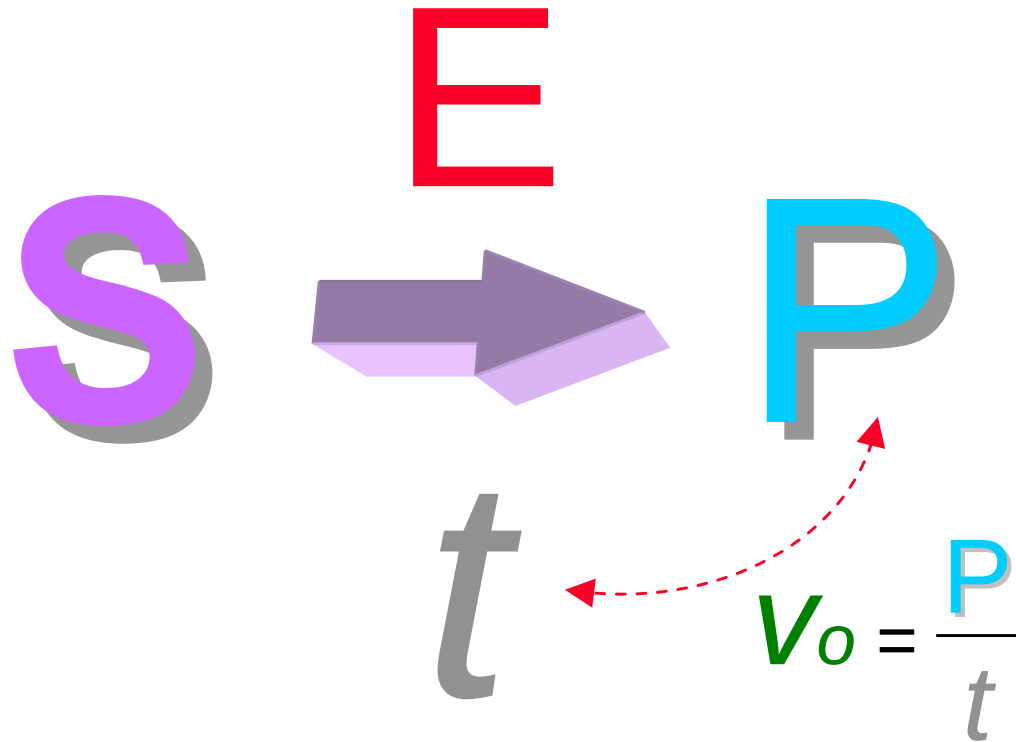


■ 酵素活性的測定

酵素量要適中

基質
要過量

$10 \times K_m$



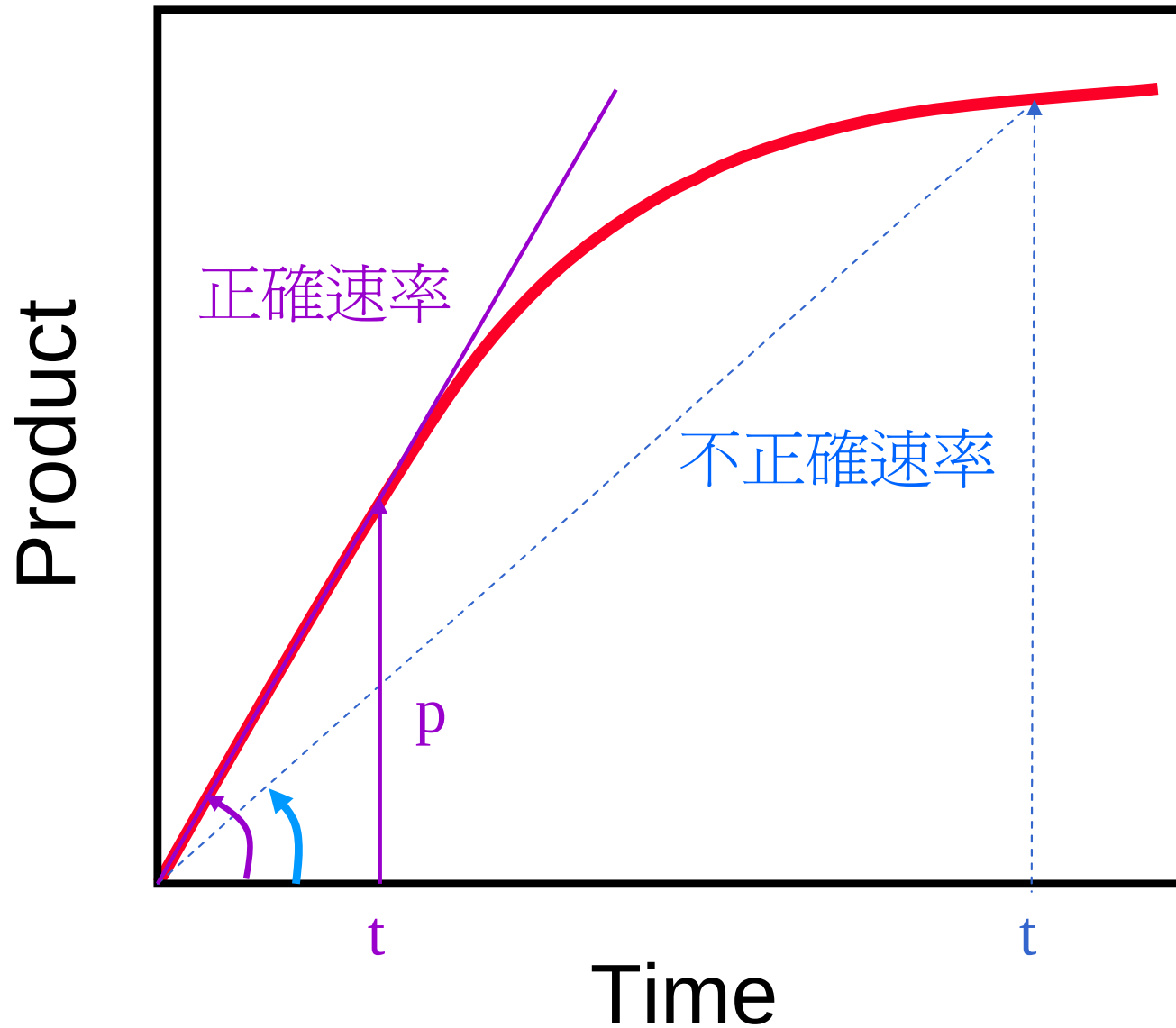
生成物
要能測得

酸鹼度

反應時間恰當

溫度

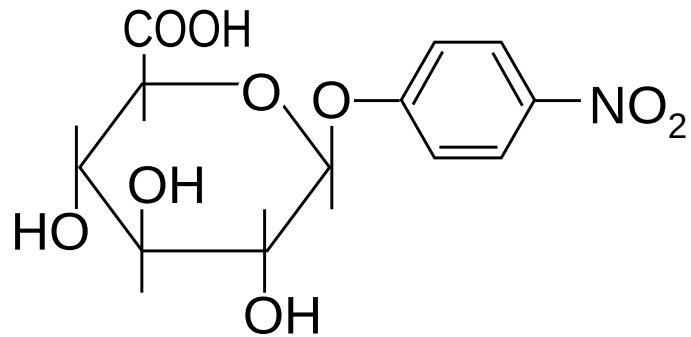
■ 測定時間內反應速率需成線性



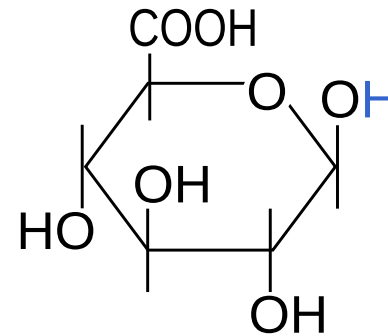
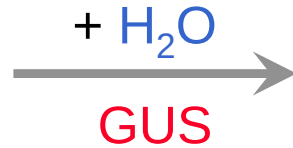
$$v = \frac{p}{t}$$

GUS 酵素活性的測定

反應物

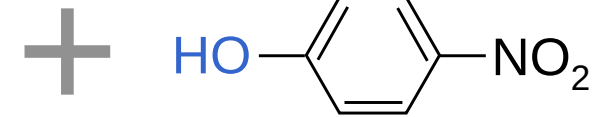


p-Nitrophenyl β -D-glucuronide
(pNPG)



β -D-Glucuronic acid

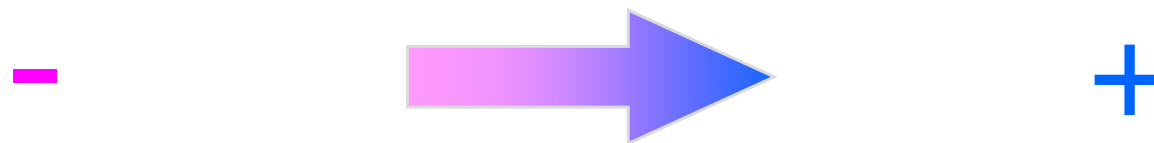
生成物



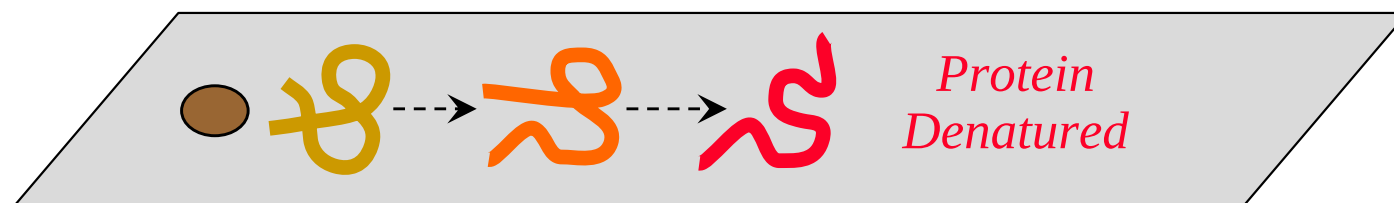
p-Nitrophenol
(yellow)

415 nm

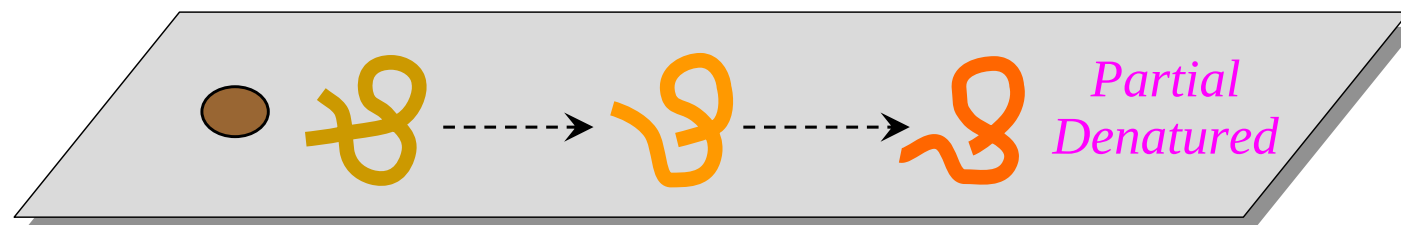
電泳形式的演進



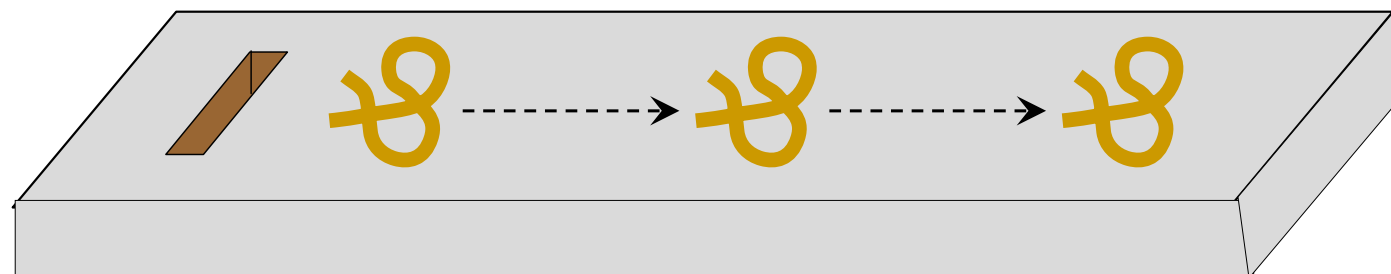
濾紙電泳 Cellulose



薄層電泳 (TLE) Cellulose acetate

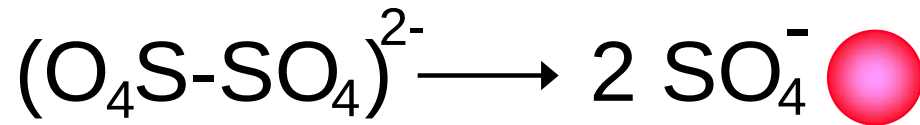


膠體電泳 Starch → Gel



■ 膠體的聚合反應

Ammonium persulfate (free radical initiator)



自由基的生成者

Acrylamide (monomer)

成膠的基本單位

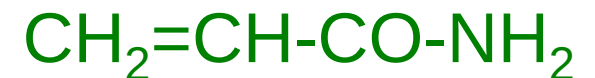
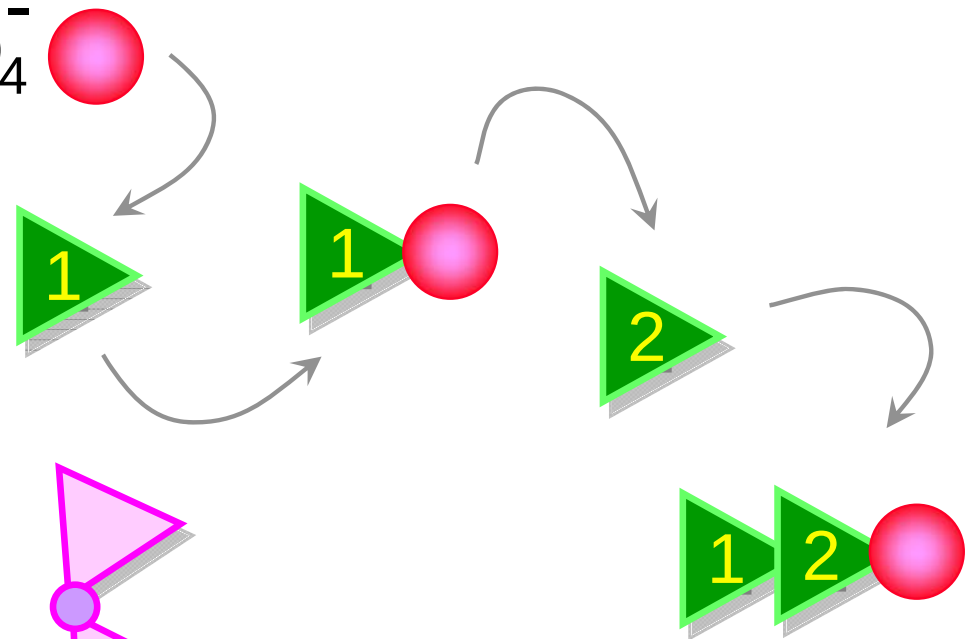
Bis(acrylamide) (bridge)

架橋使聚合產生分枝

TEMED (catalyst)

幫助傳遞自由基的催化劑

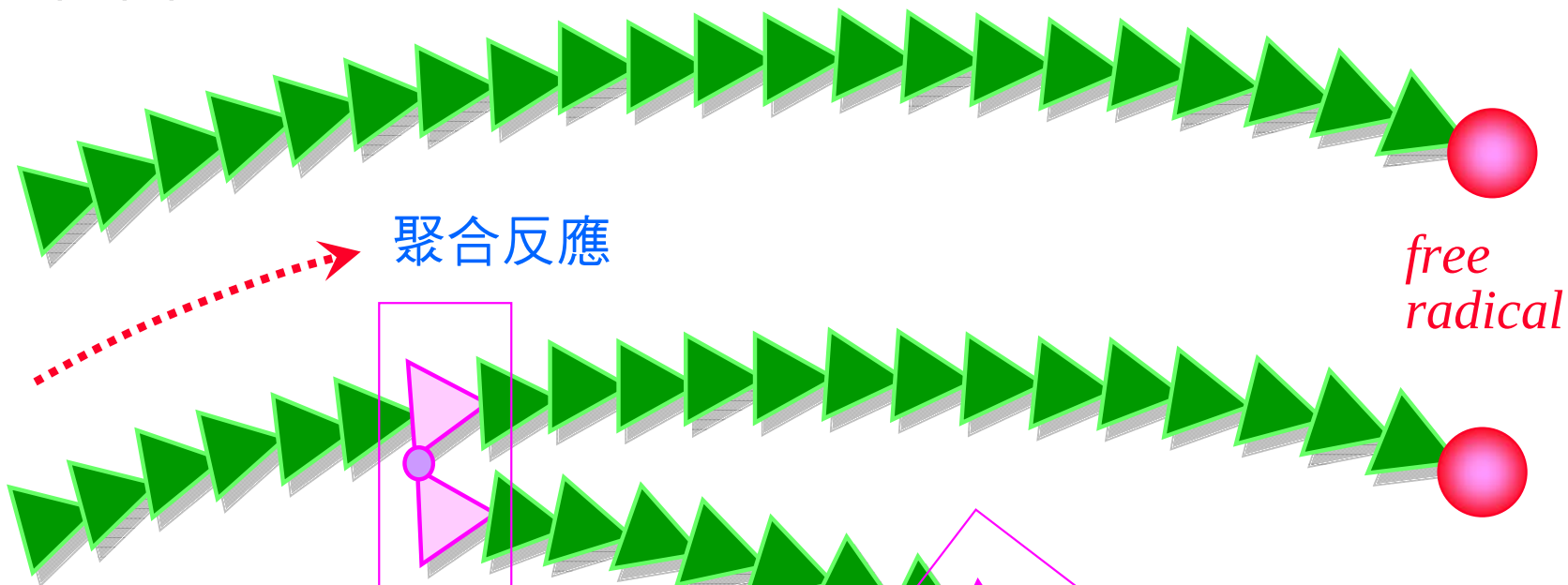
SDS (Sodium dodecyl sulfate)



Acrylamide 有毒性！
澱粉加高熱會誘生？

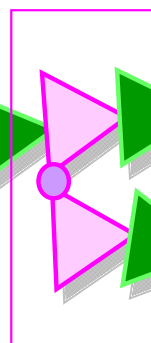
單體聚合反應

鑄膠反應



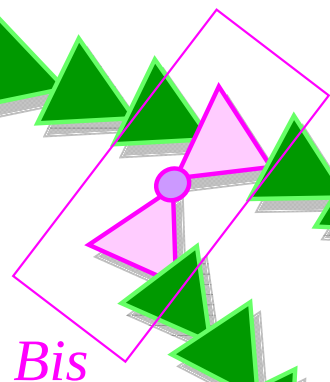
聚合反應

free radical



Bis

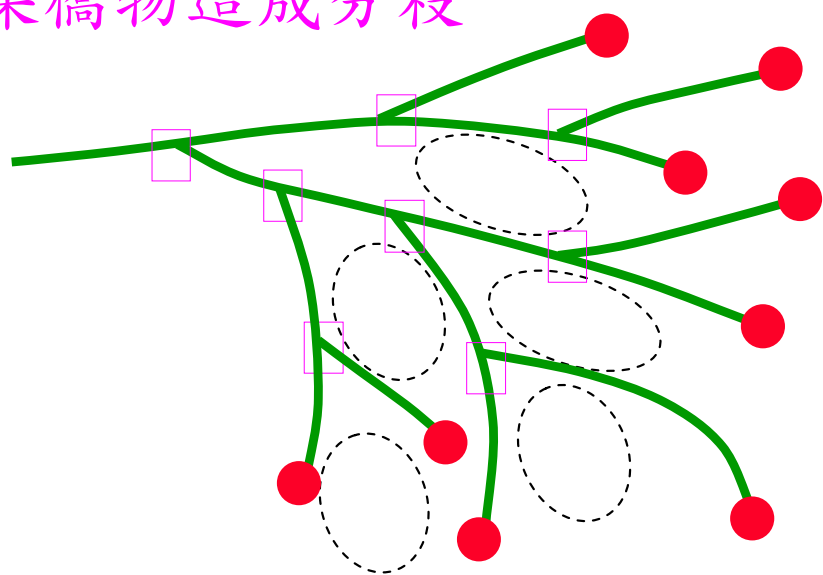
交錯連結



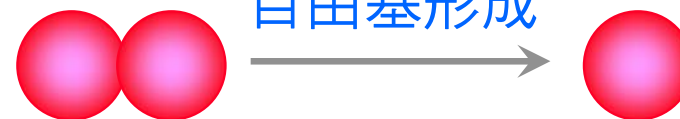
Bis

端點自由基
可再延續

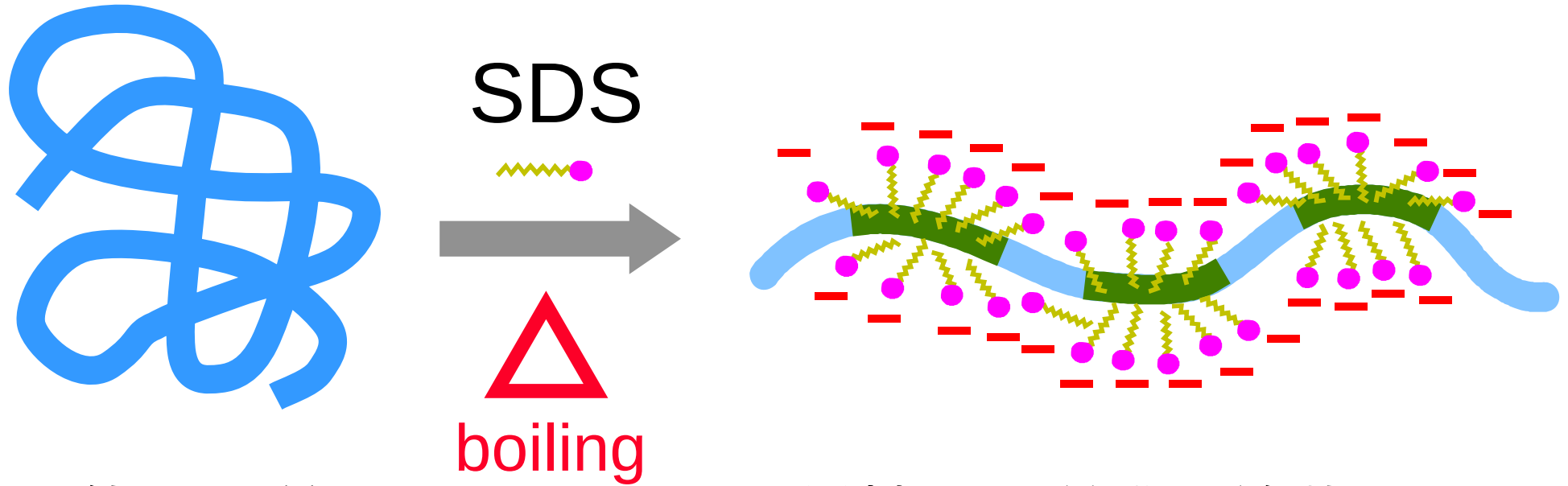
架橋物造成分枝



自由基形成



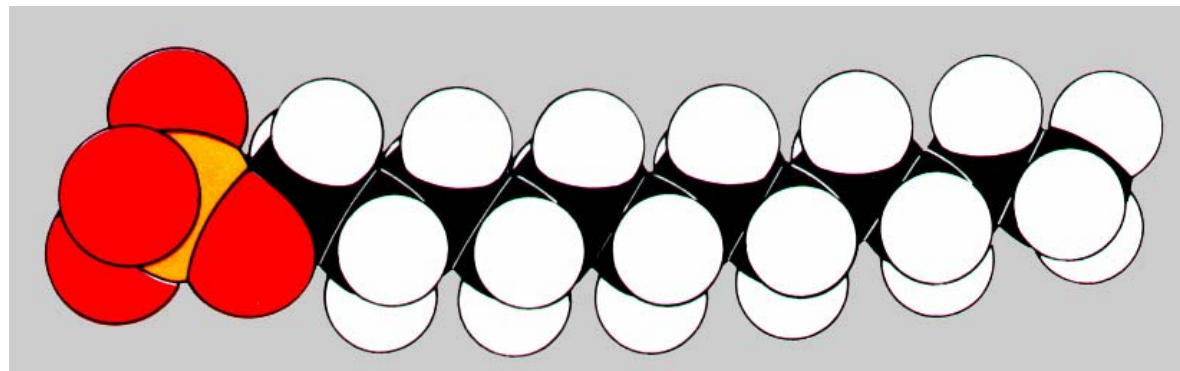
■ SDS 在蛋白質表面 均勻敷上 一層負電



原態蛋白質

變性蛋白質成一線狀分子
並且均勻帶上一層負電荷

極性頭部



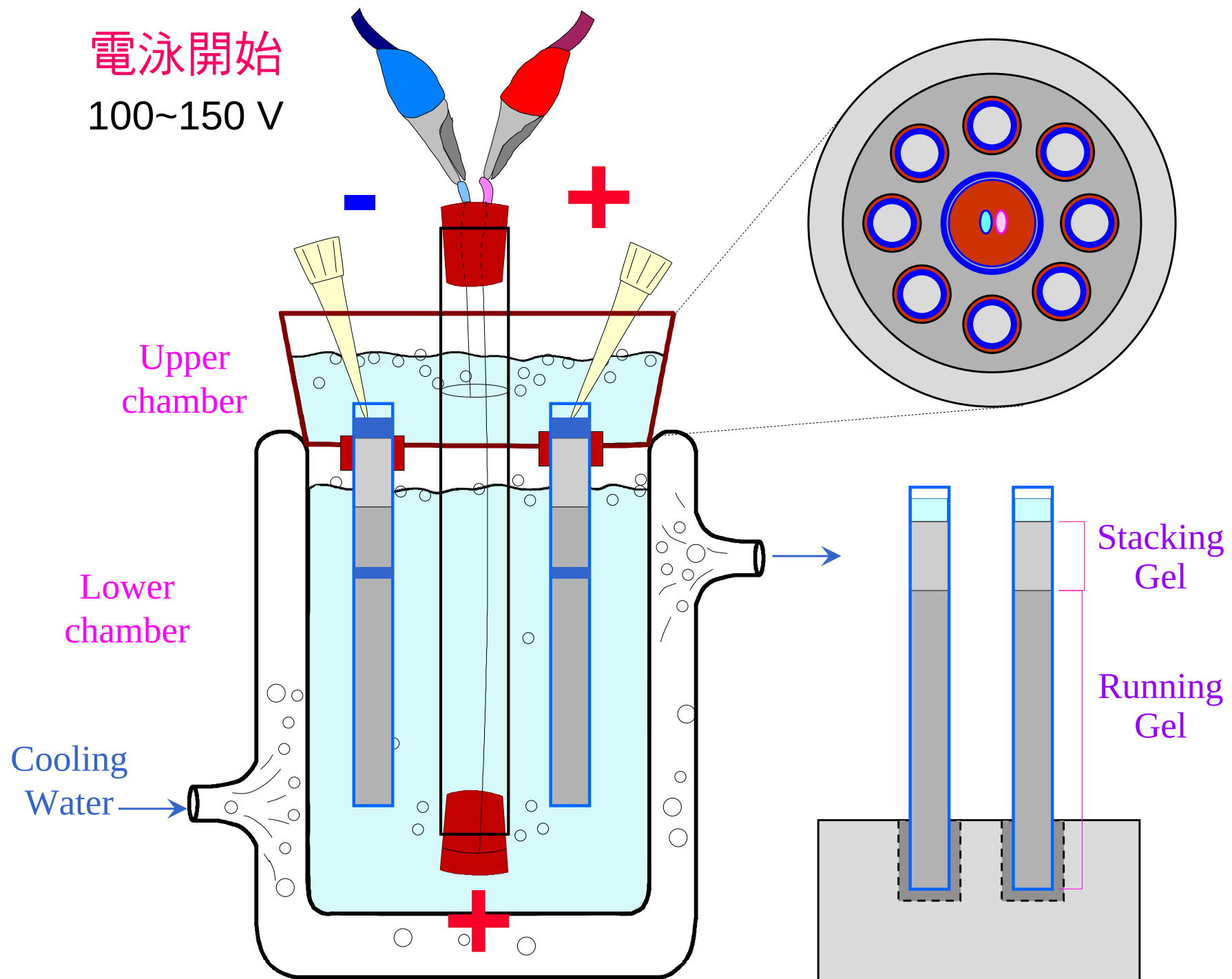
非極性尾部

■ 電泳膠體系統的組成

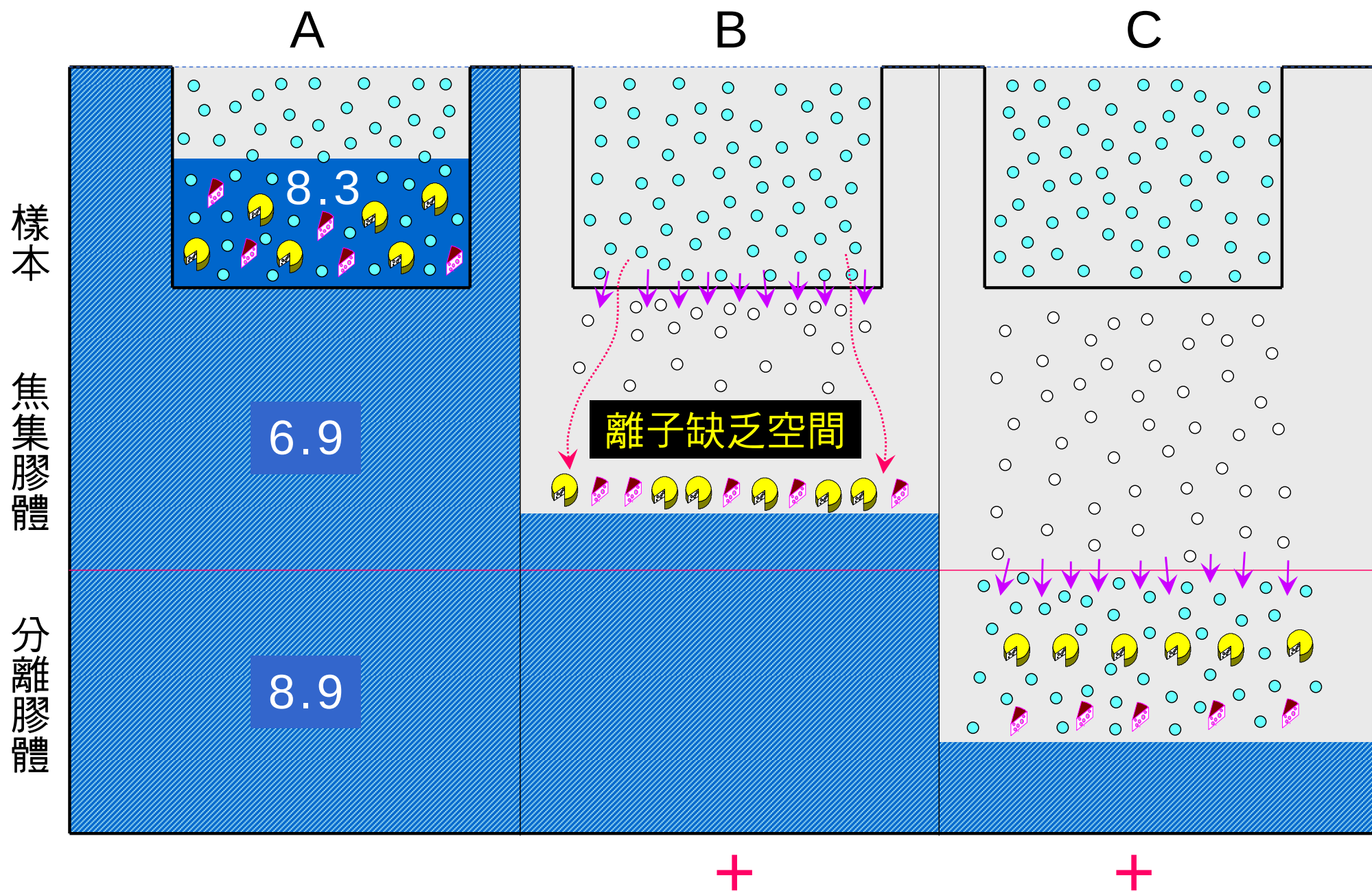
電泳系統		緩衝液	pH	膠體濃度	
1	上層 (負極) 緩衝液		Tris-glycine	8.3	-
2	樣本溶液		Tris-glycine	8.3	-
3	膠體	聚焦膠體	Tris-HCl	6.9	5%
4		分離膠體	Tris-HCl	8.3	7.5~20%
5	下層 (正極) 緩衝液		Tris-glycine	8.3	-

● 膠體不連續性有聚焦樣本的作用

■ 直立式電泳裝置



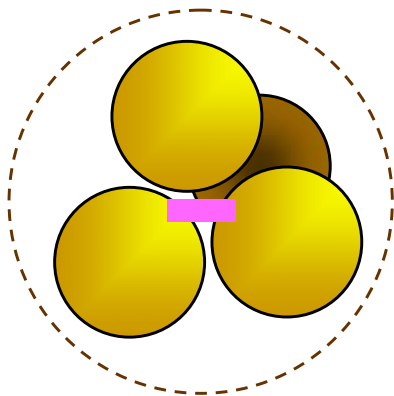
■ 焦集膠體對蛋白質分子的焦集作用



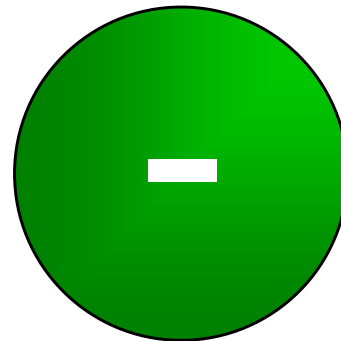
■ 三種不同性質蛋白質的電泳比較

Protein	Quaternary Structure	Molecular Weight	pI	Mobility	
				Native PAGE	SDS-PAGE
X	Tetramer	(40,000)x4	5.8	Slow	Fast
Y	Monomer	88,000	5.2	Fast	Slow
Z	Monomer	60,000	9.3	Upward	Medium

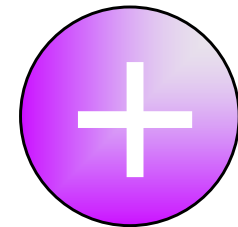
X



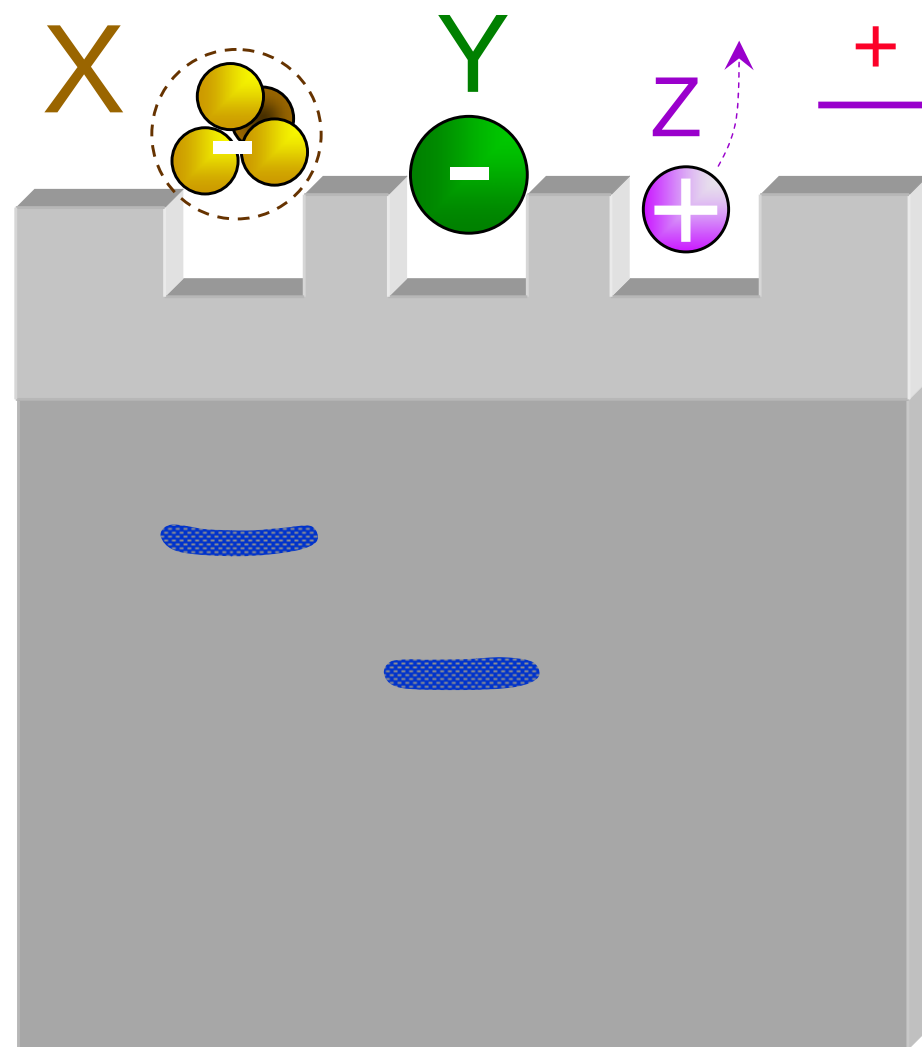
Y



Z



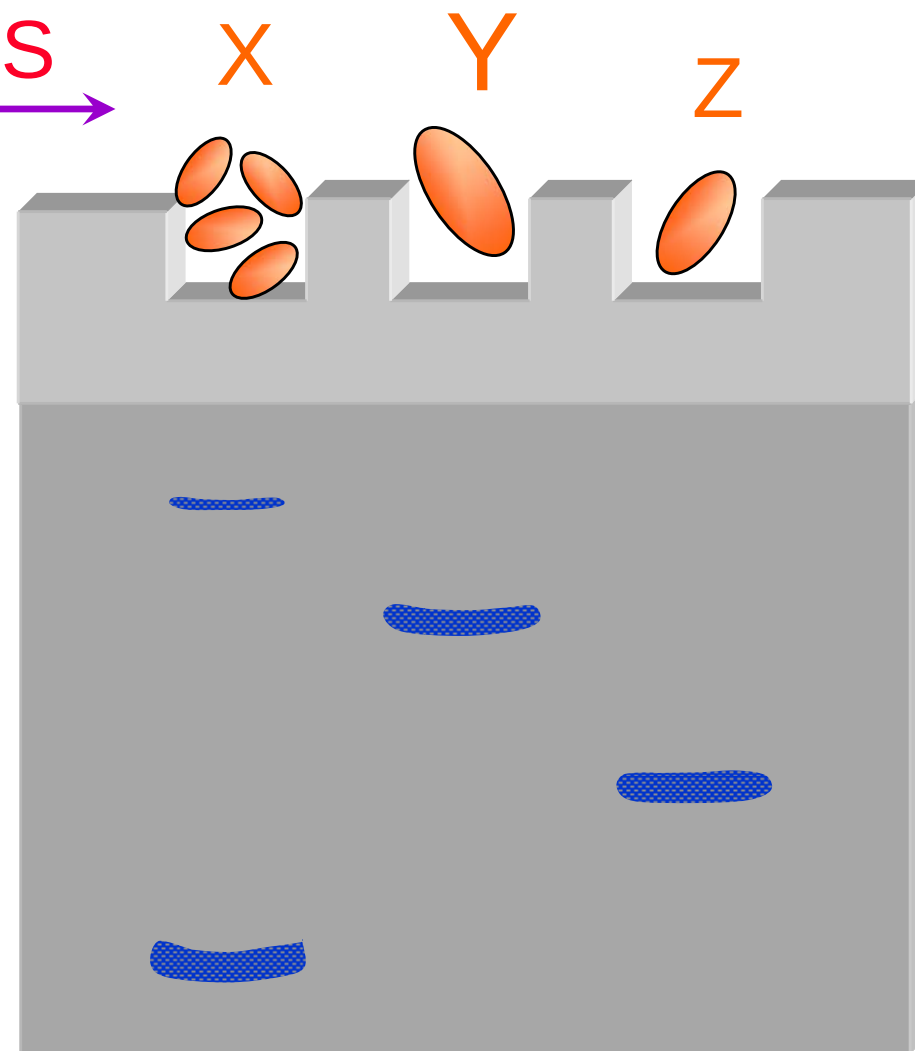
Native-PAGE



+

分子量 及 淨電荷密度
均影響泳動率

SDS-PAGE

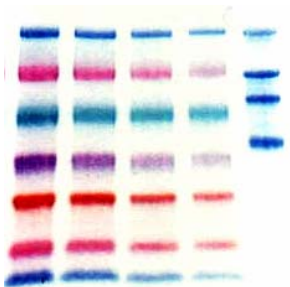
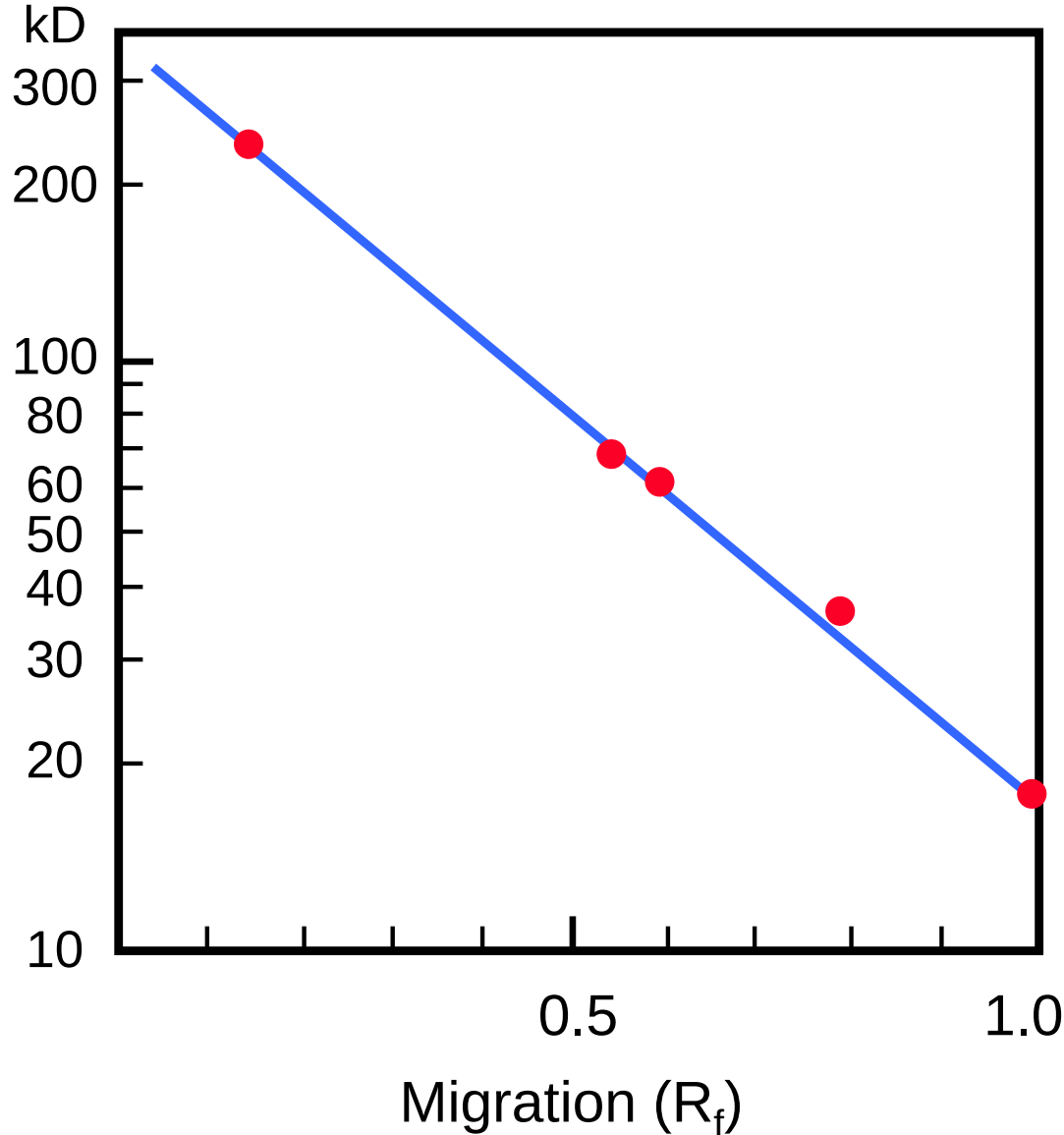


+

只有 分子量 影響泳動率

■ 單元體分子量的測定： SDS-PAGE

Mol mass



kD

330
220

67
60

36

18.5

kD

94

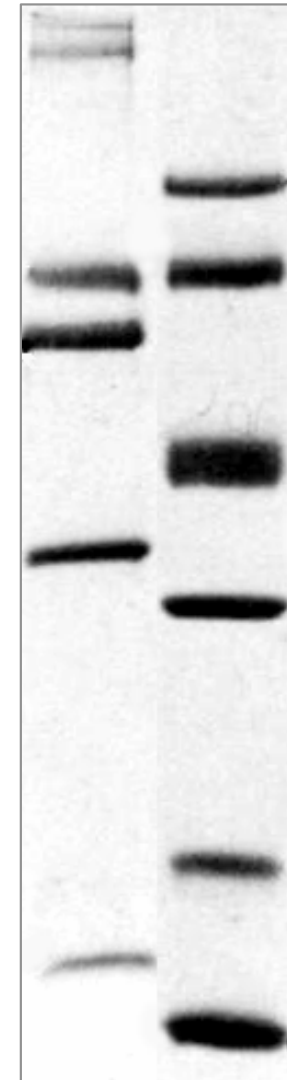
67

43

30

20.1

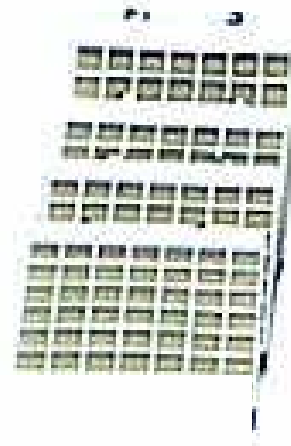
14.4



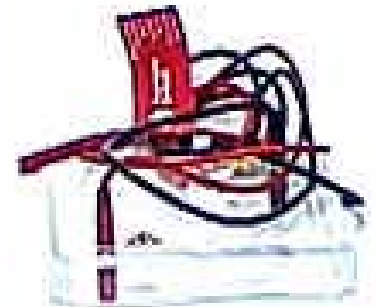
■ 電泳槽及相關設備



轉印三明治



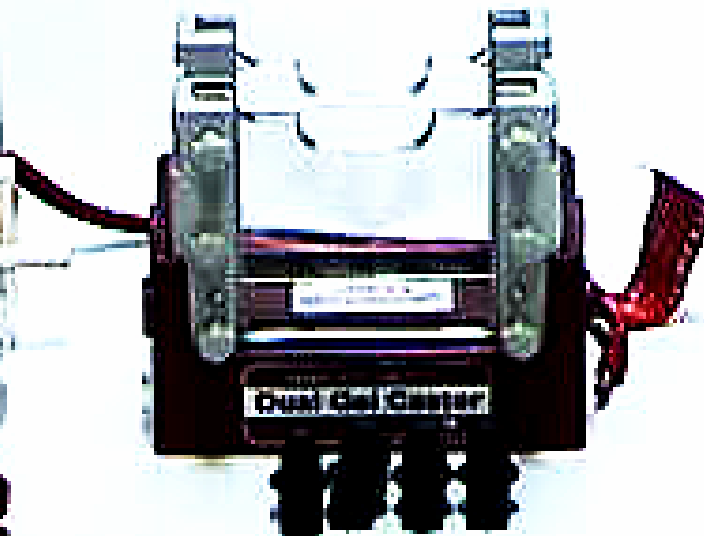
轉印槽



電泳槽



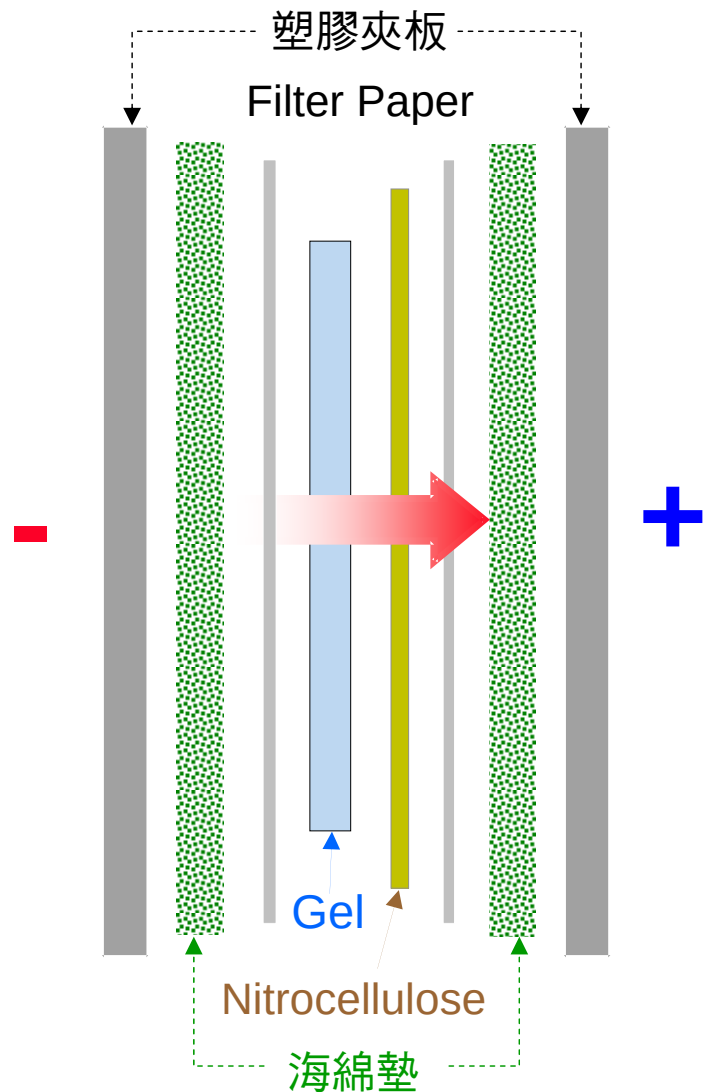
鑄膠器



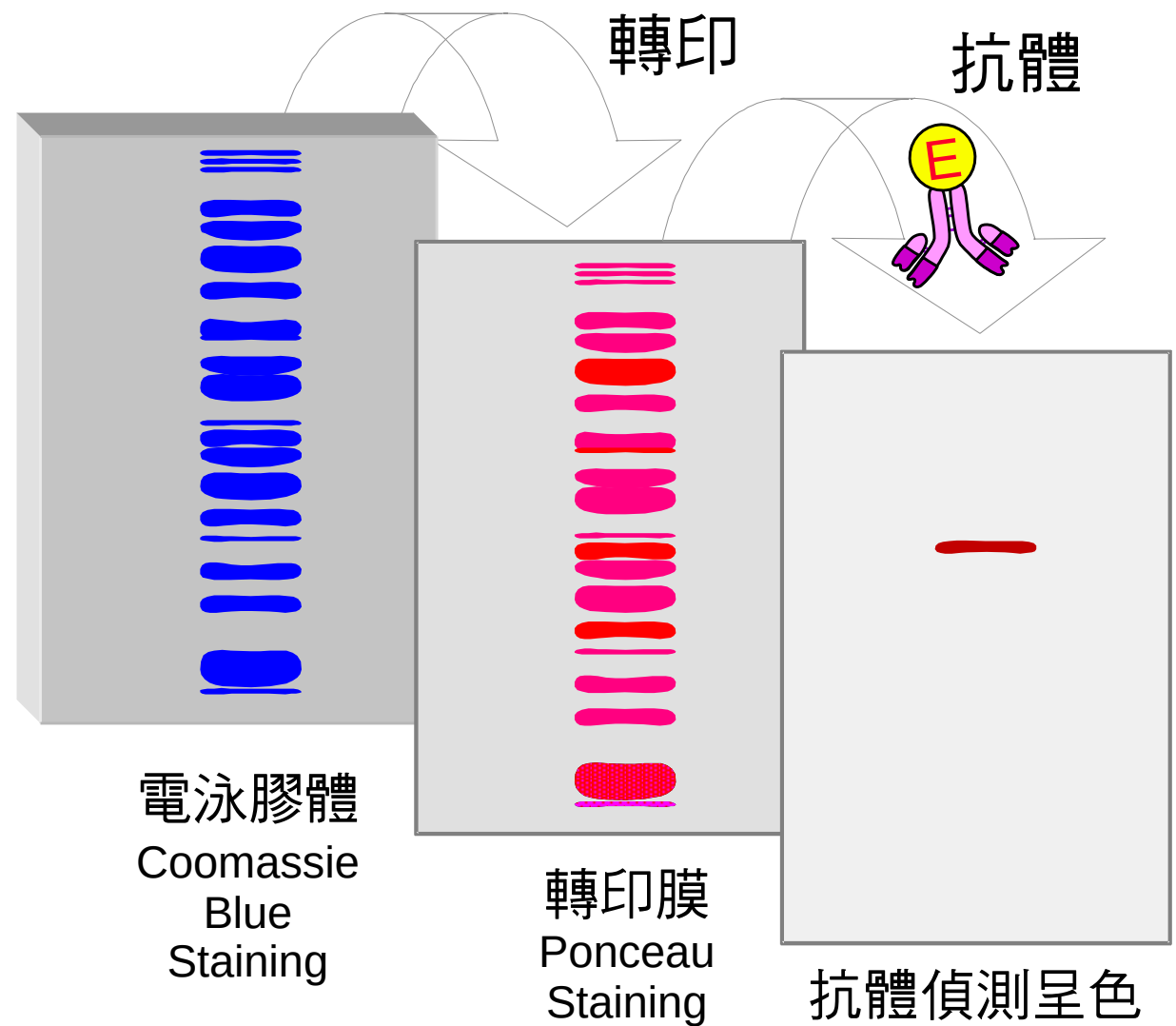
供電器

轉印及免疫染色流程

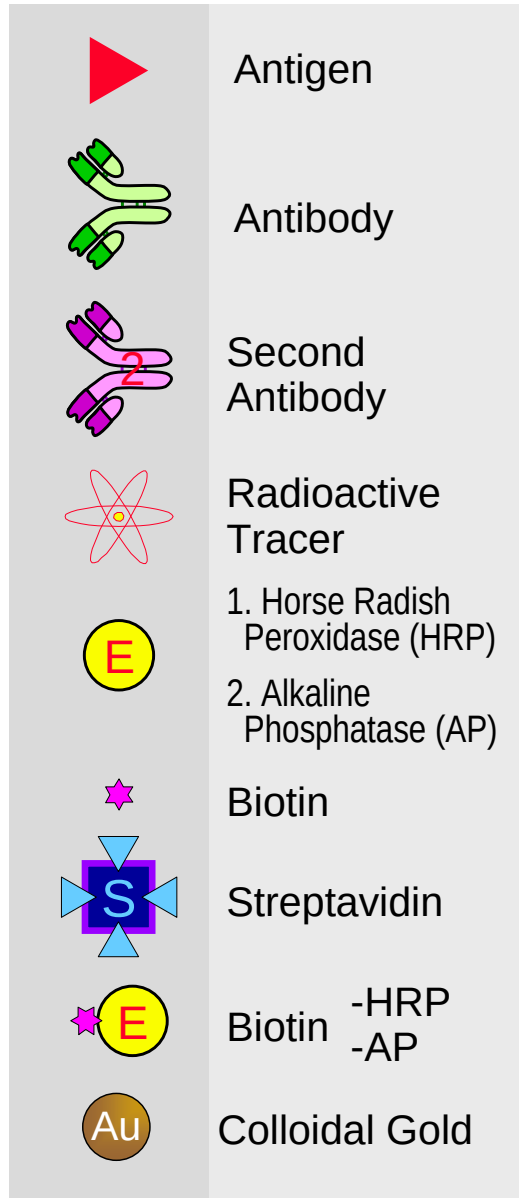
A 轉印三明治



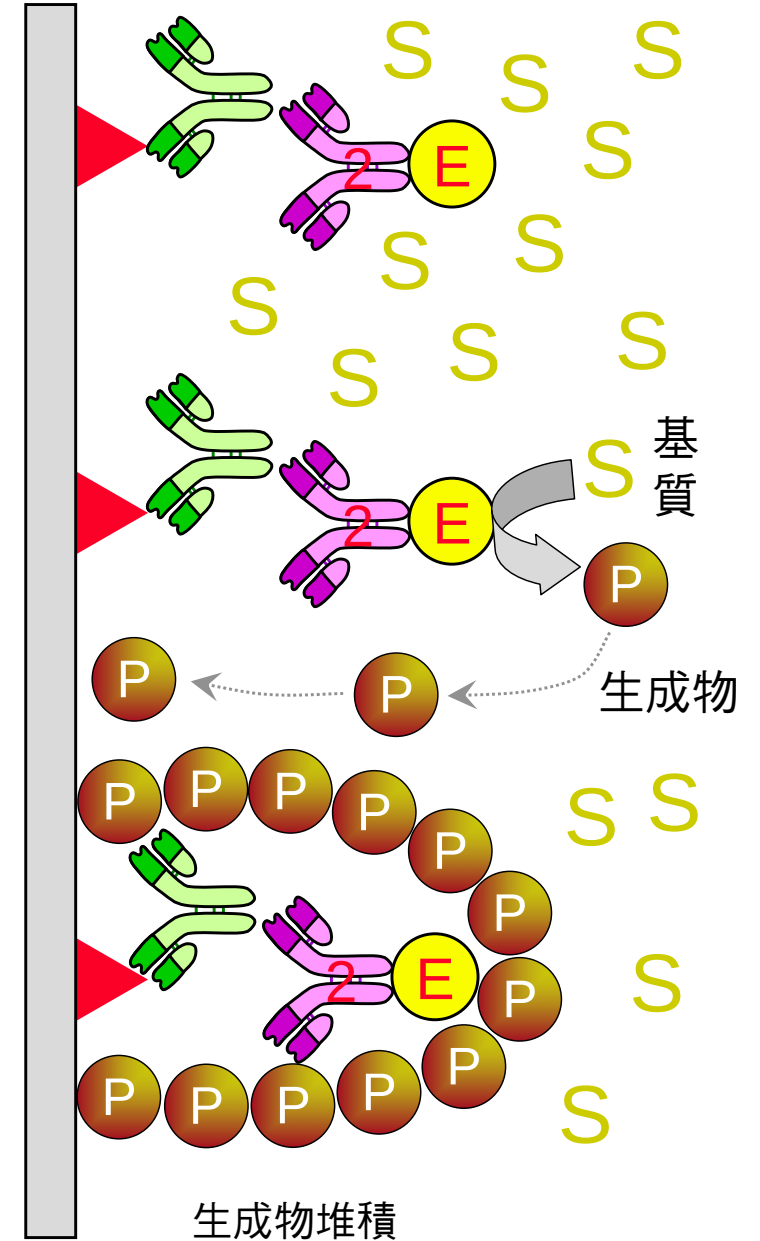
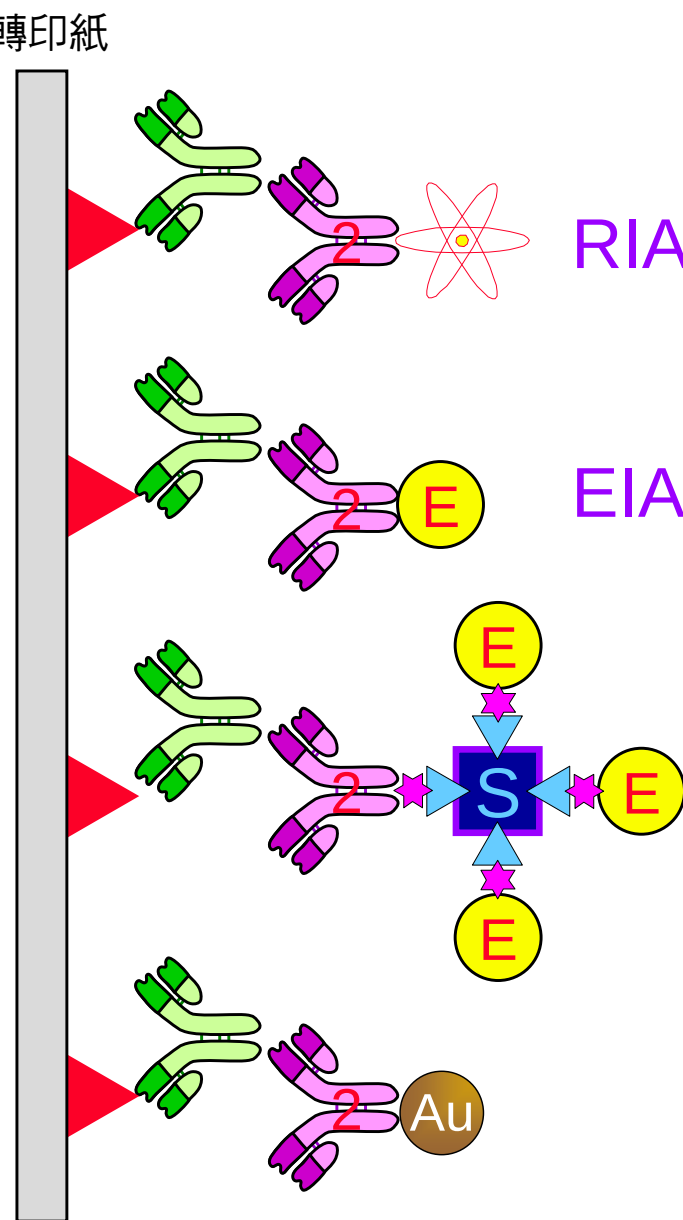
B 免疫染色流程及結果



免疫轉印的種類與呈色機制

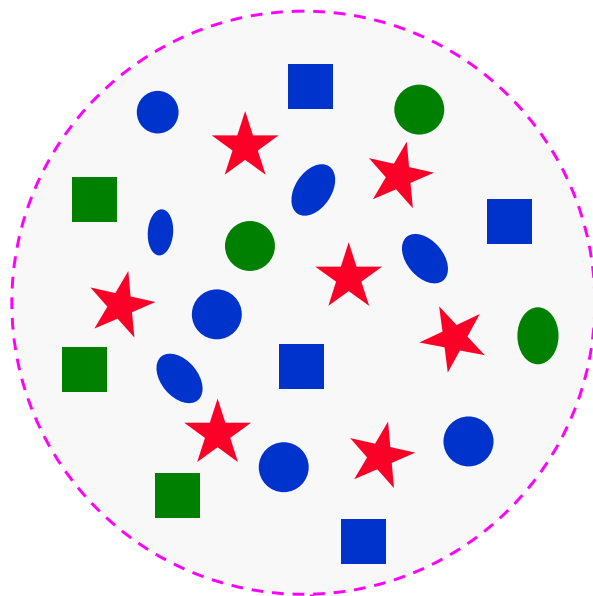
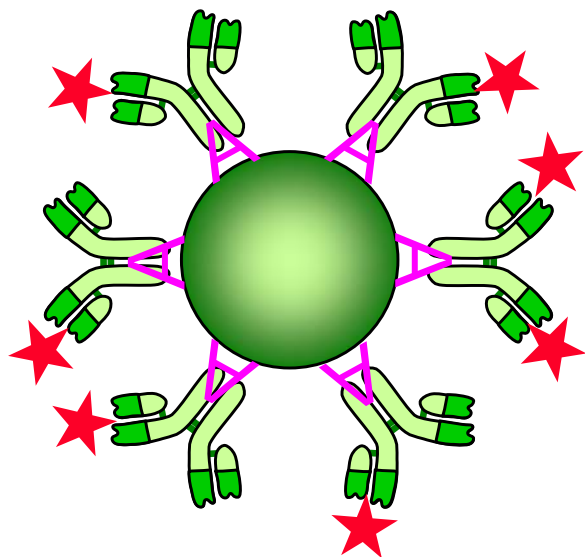


轉印紙



擔體免疫沈澱的原理及應用

擔體免疫沈澱

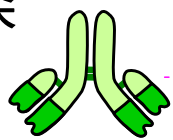


細胞粗抽取液

離心分離後
以 SDS-PAGE 電泳檢定

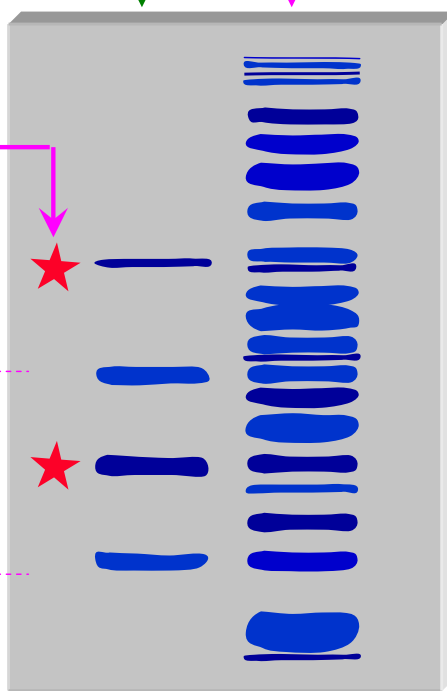
抗原可能有
兩個次體

抗原有輕鏈及
重鏈各兩條



H

L



■ 澱粉磷解酶染色方法比較

膠體過濾法各分劃

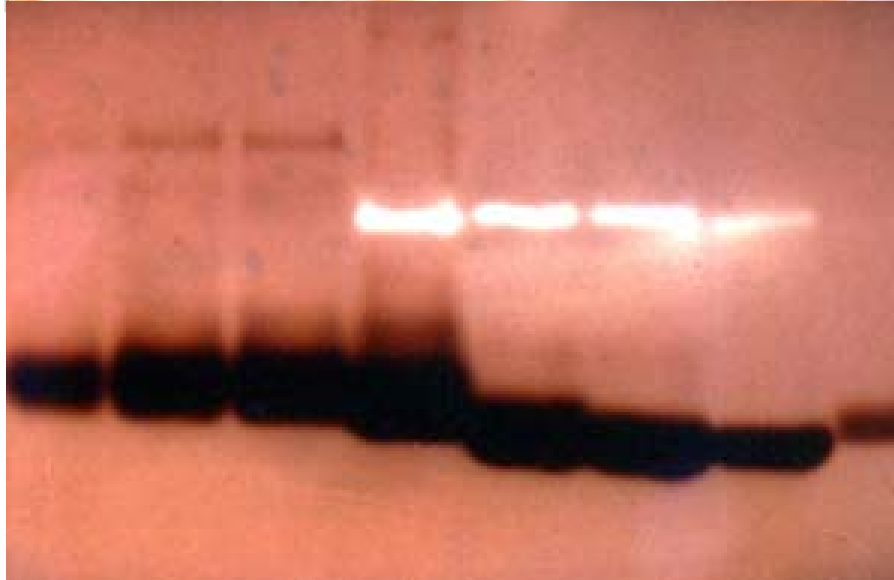
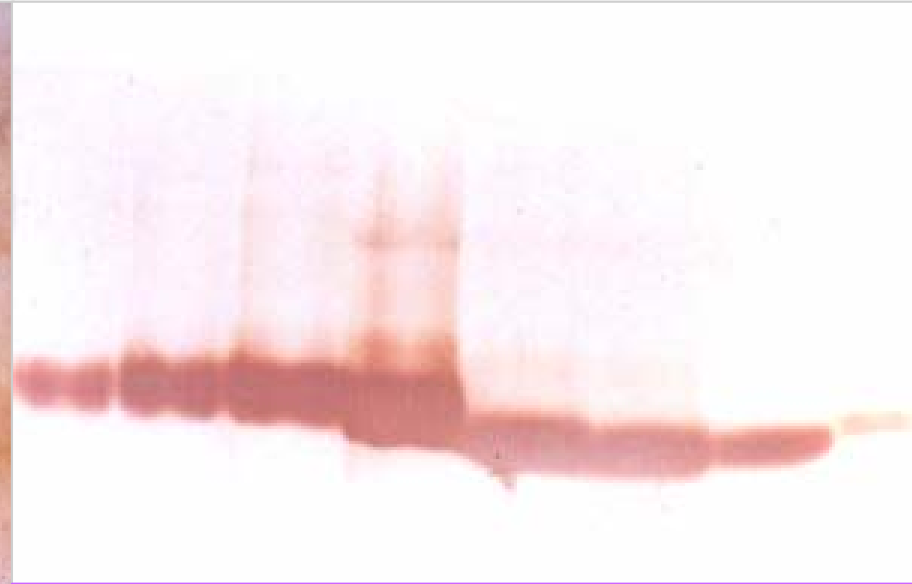
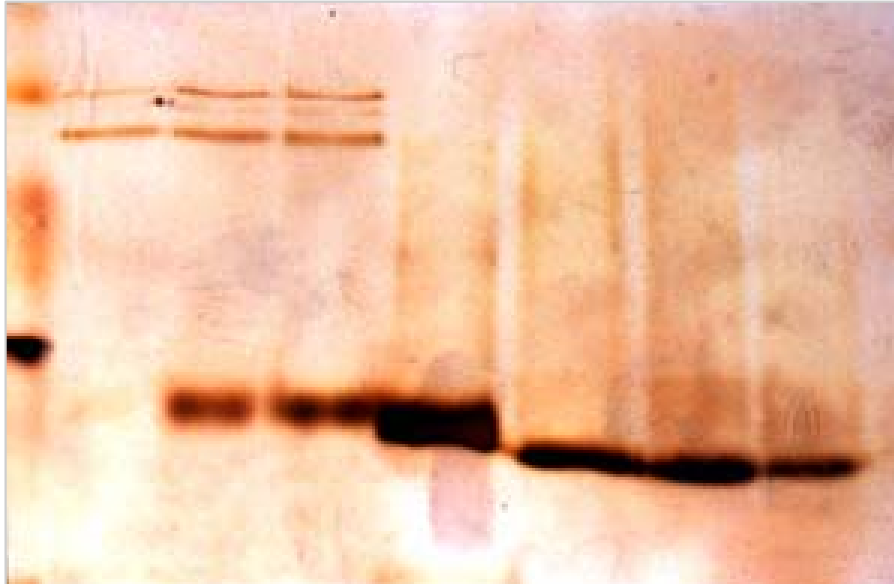
免疫轉印

硝酸銀

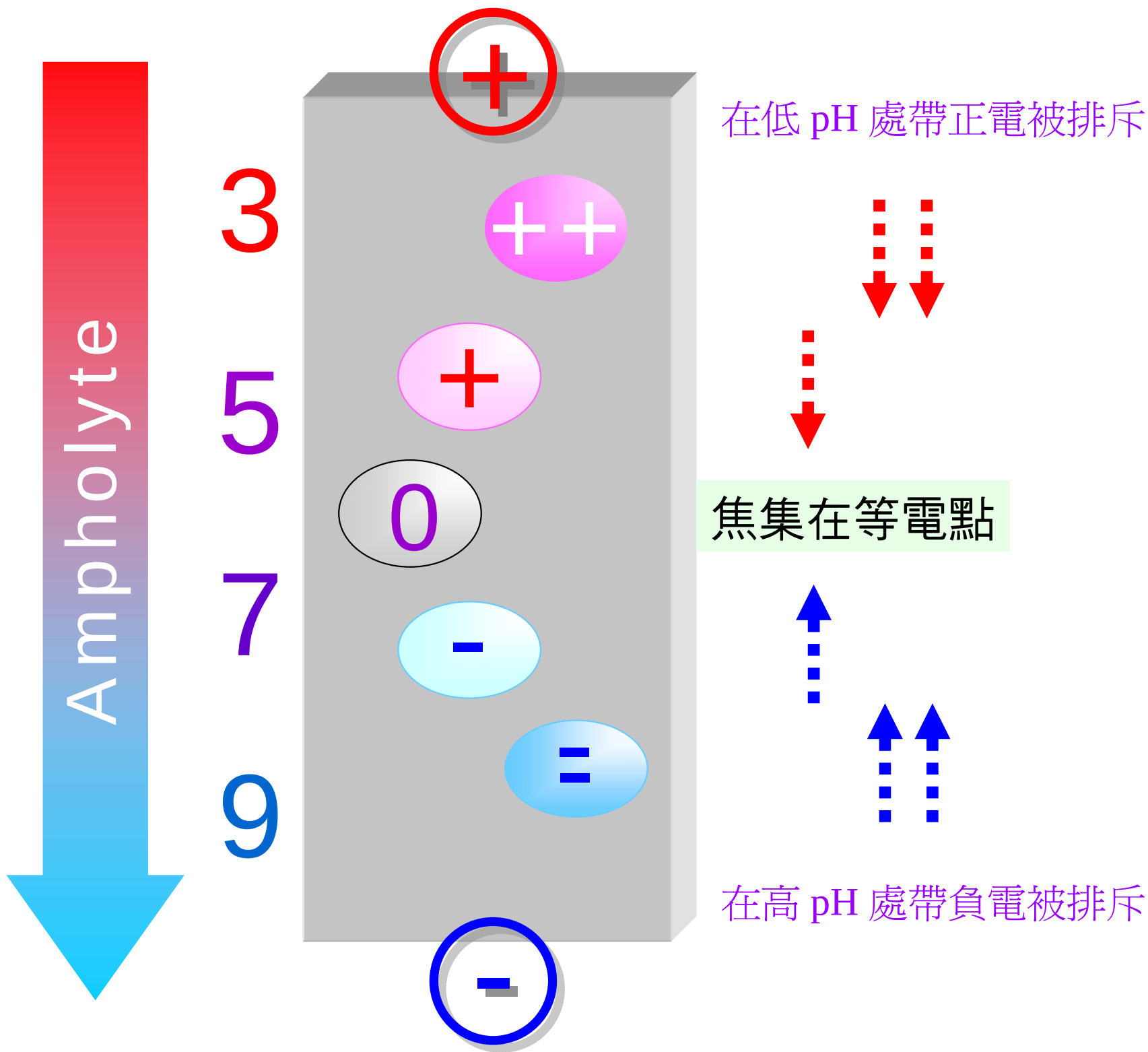
Disc-PAGE

活性染色

SDS-PAGE

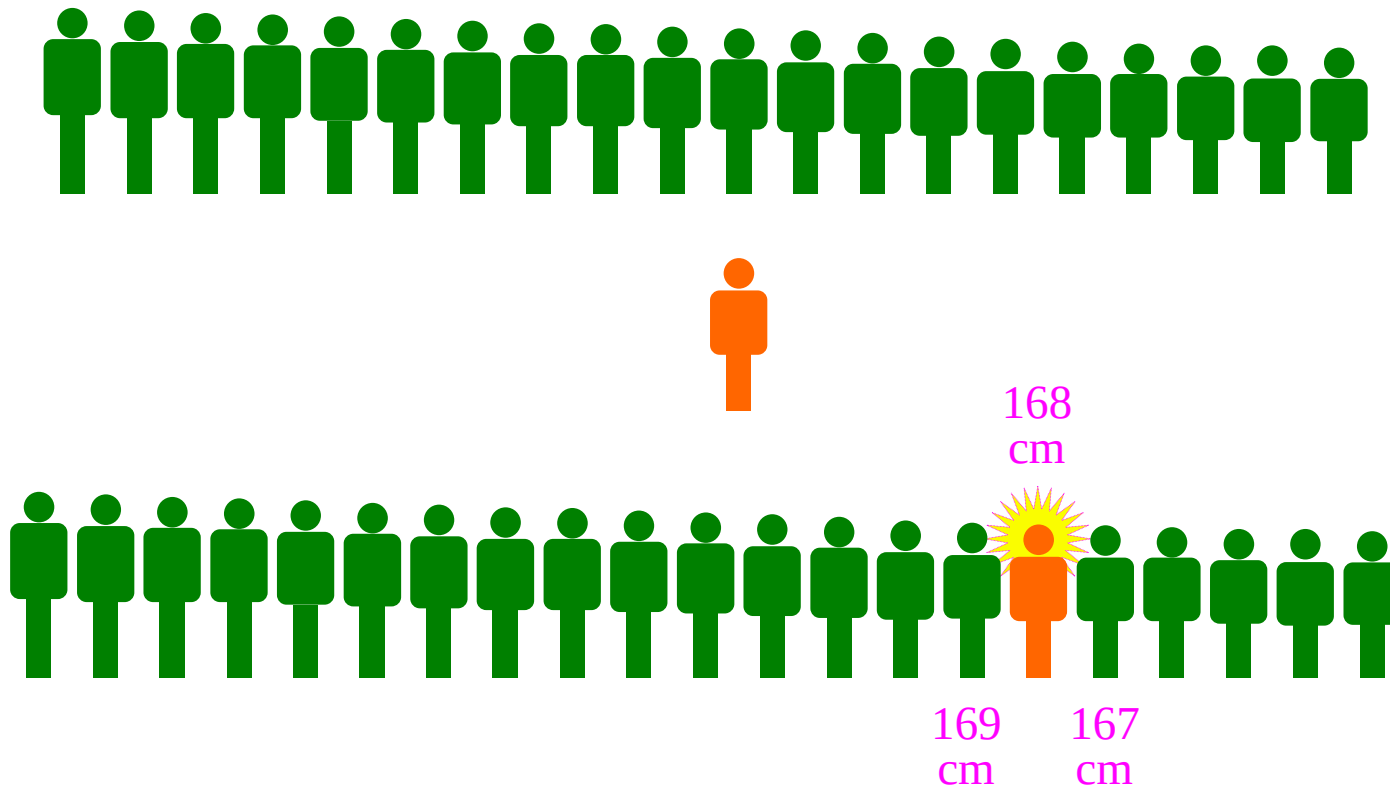


■ 等電聚焦法的聚焦方式



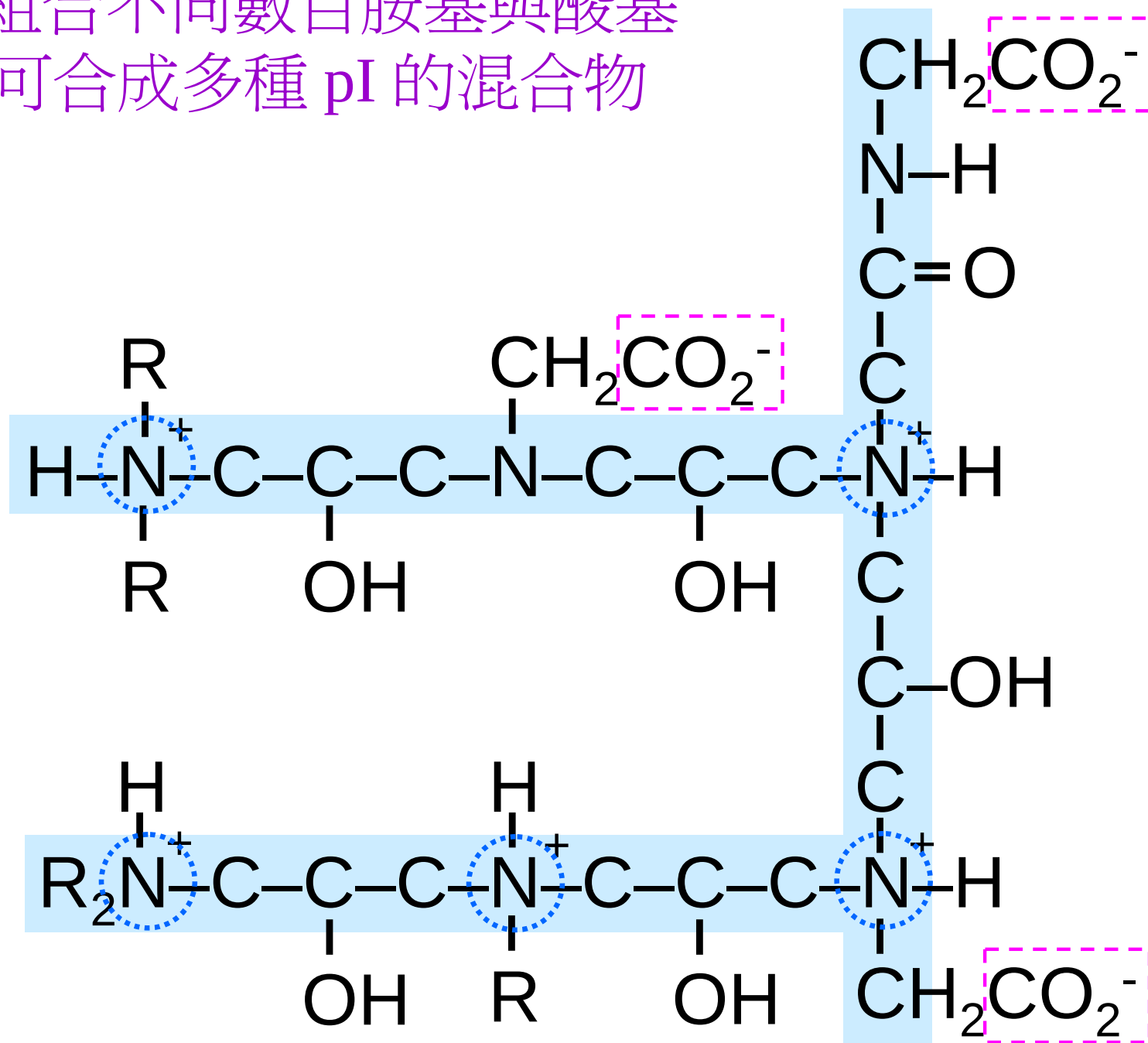
3.3.2 等電聚焦法原理

樣本分子 在一已知 pH 梯度 中聚焦

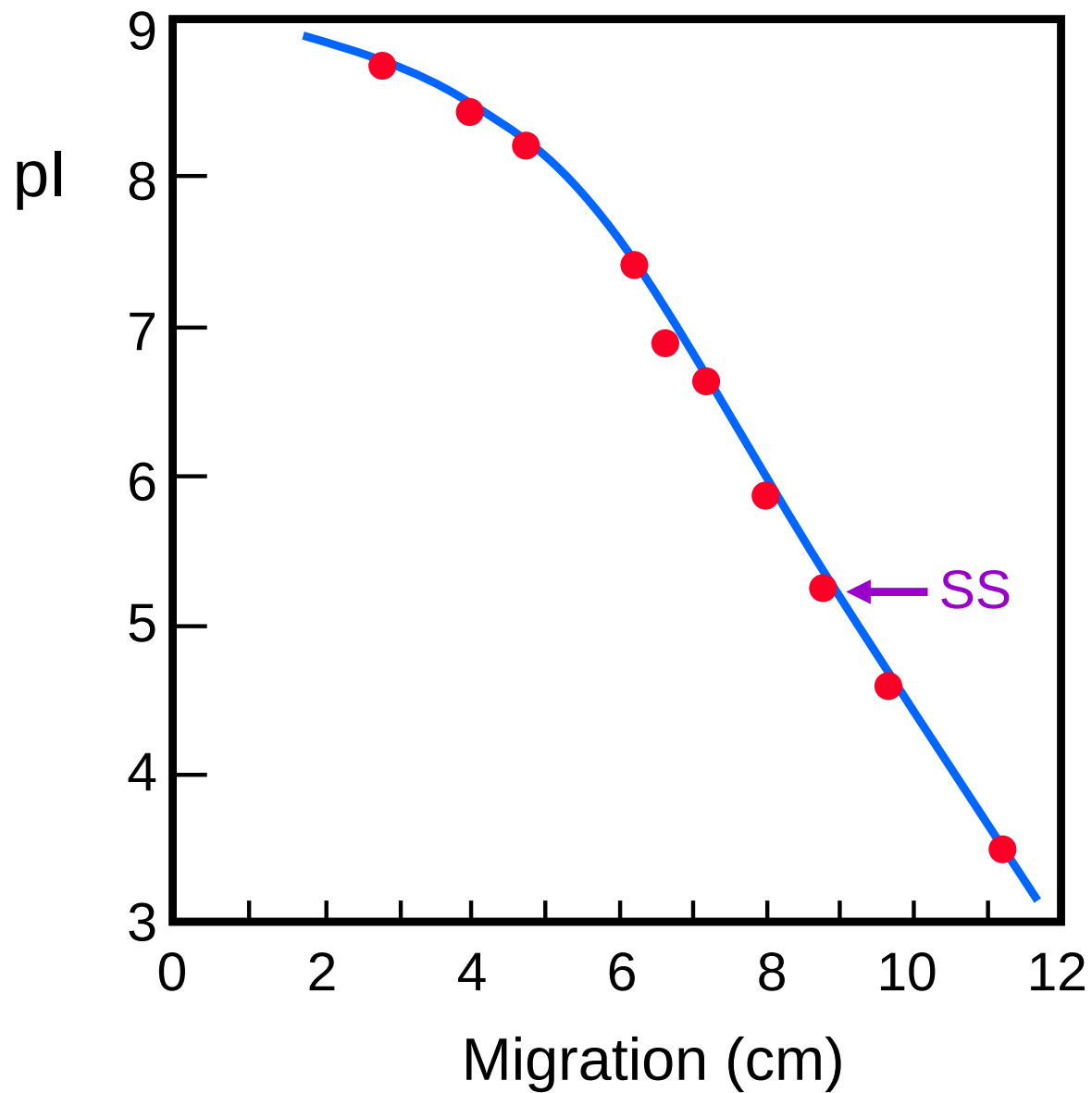


■ 等電聚焦的雙性離子構造

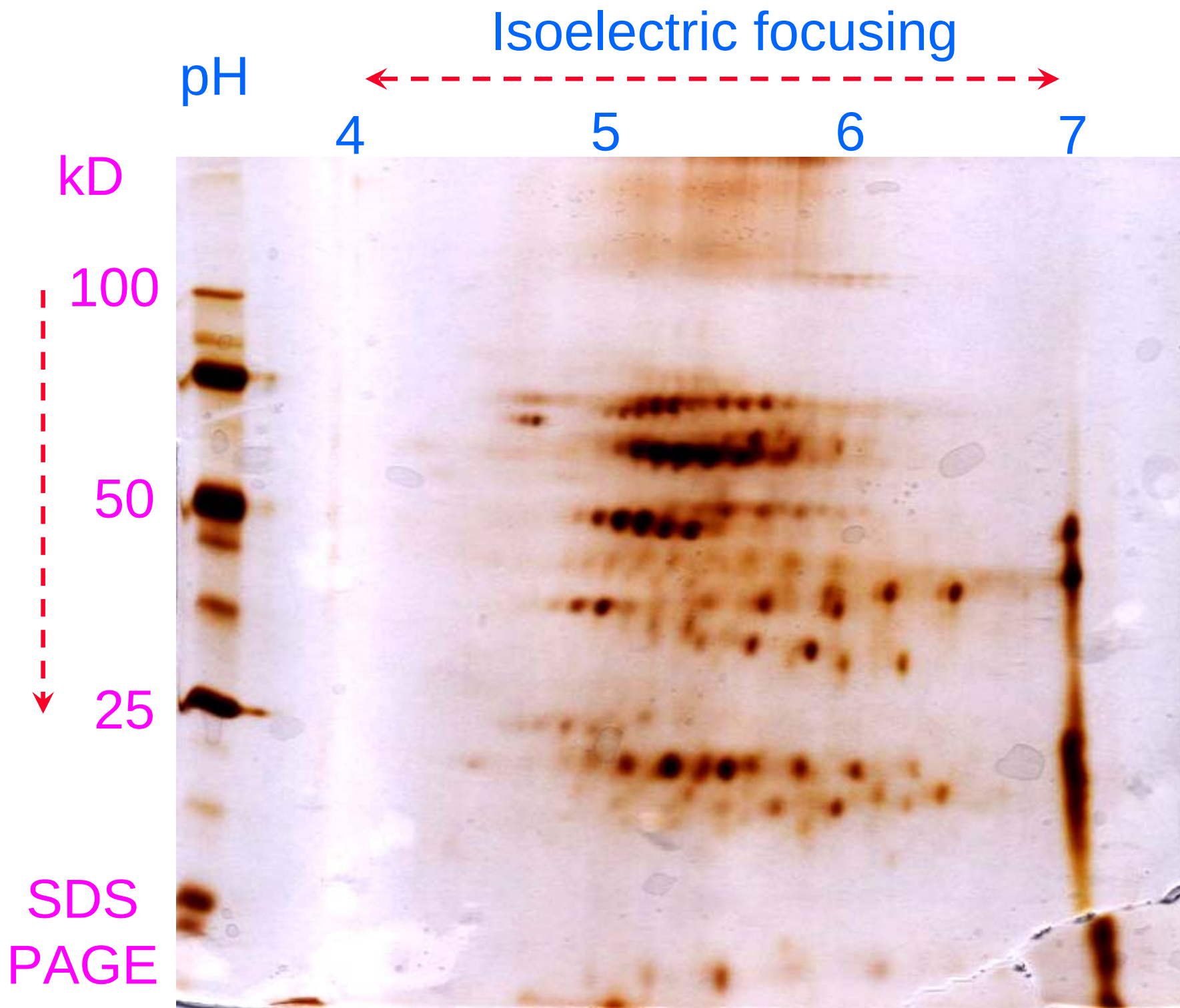
- 組合不同數目胺基與酸基
可合成多種 pI 的混合物



■ 等電點標準校正線： IEF

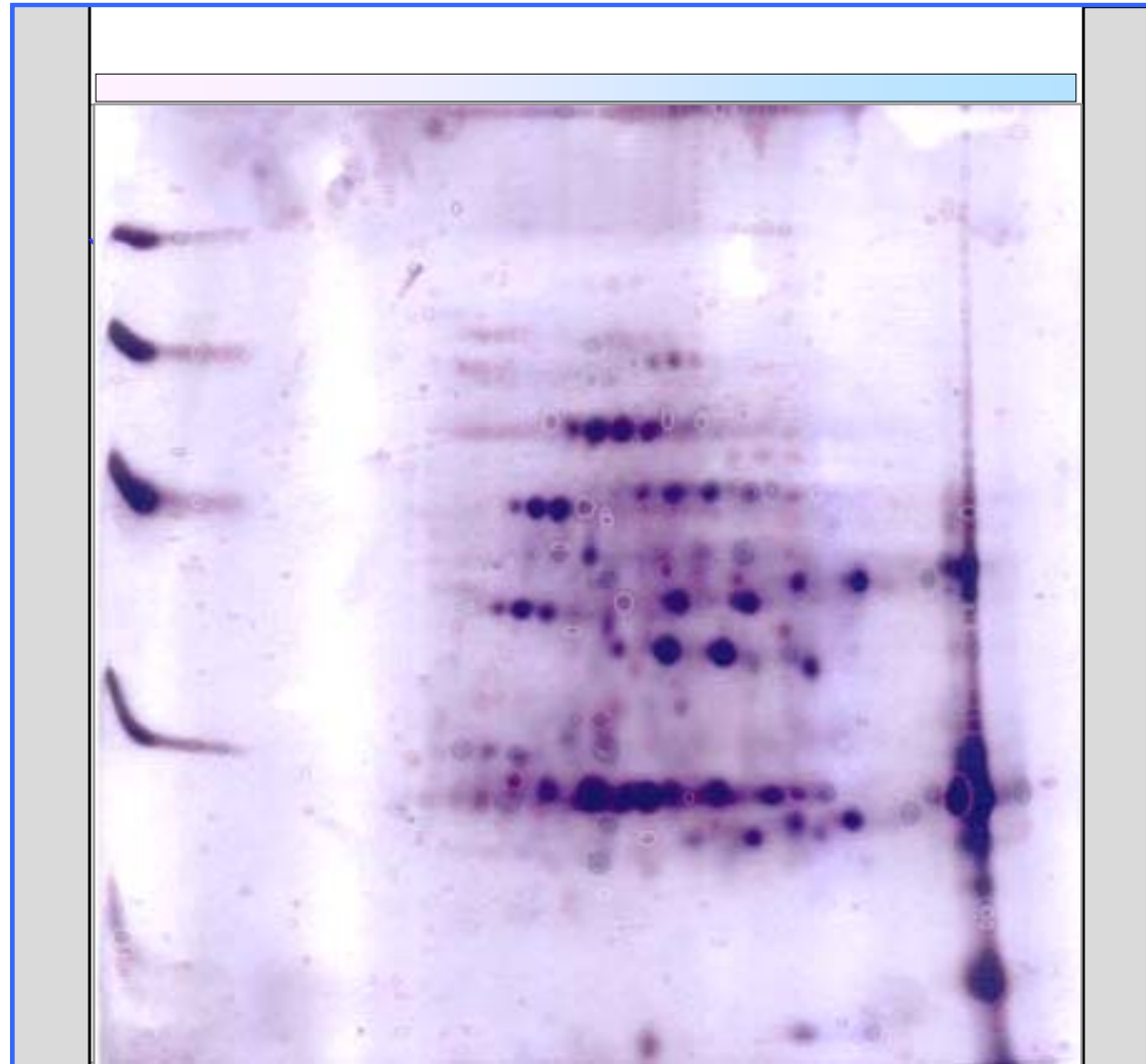


■ 二次元電泳



■ 二次元電泳操作流程

(1) IEF 等電聚焦電泳

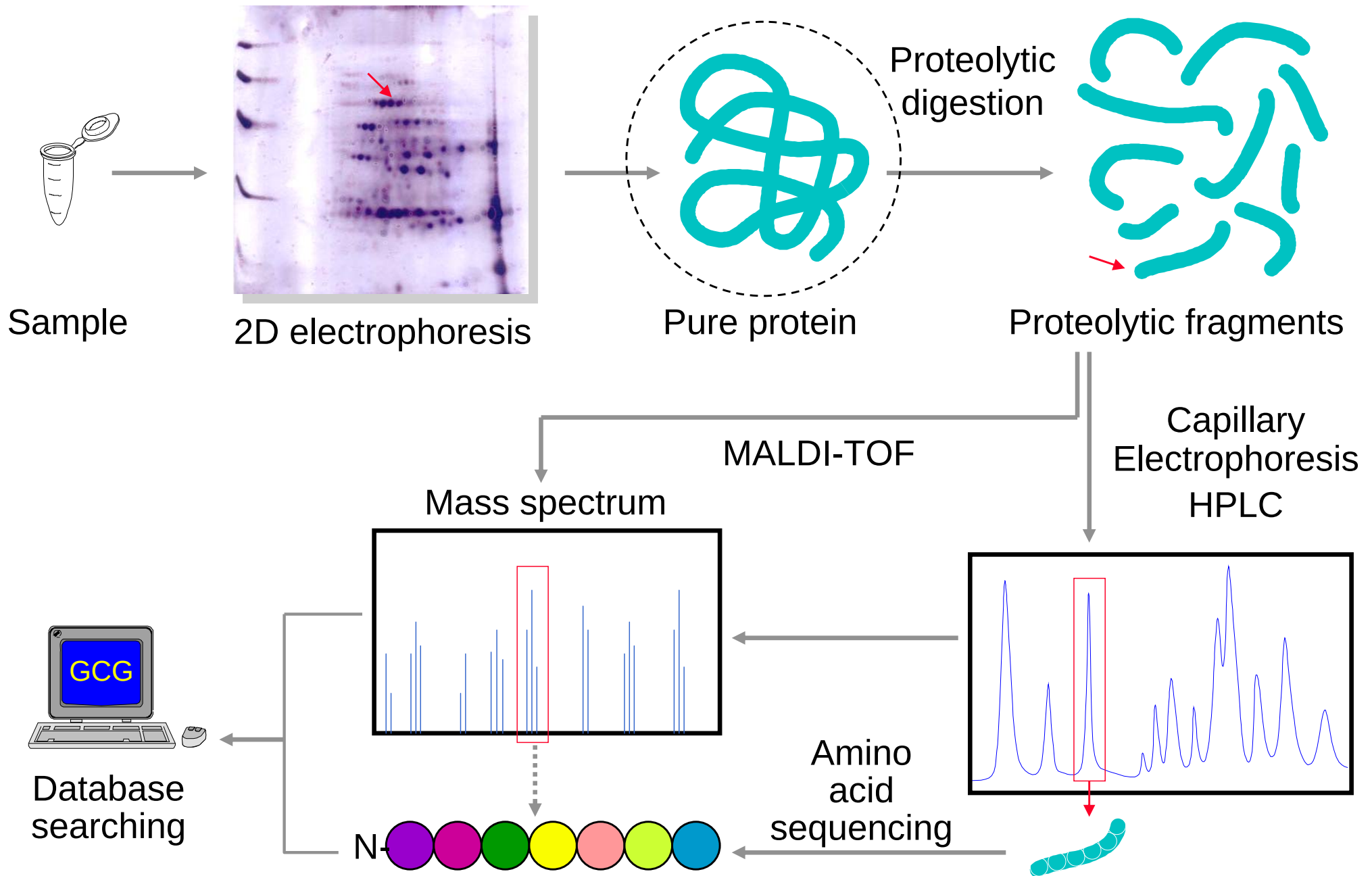


(2) SDS-PAGE 分離膠體

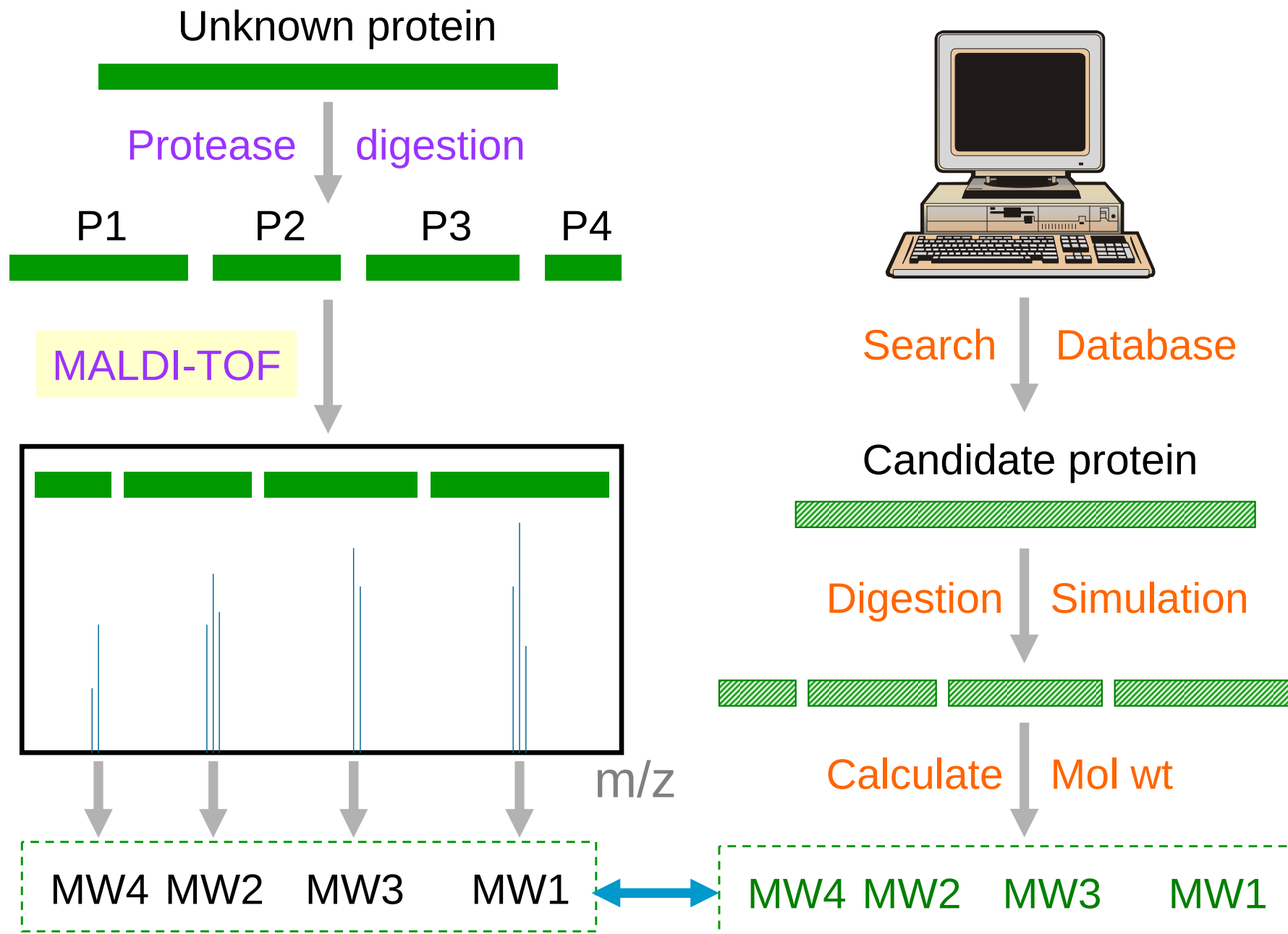
(3) 染色脫色



蛋白質體可綜觀蛋白質的消長與身分

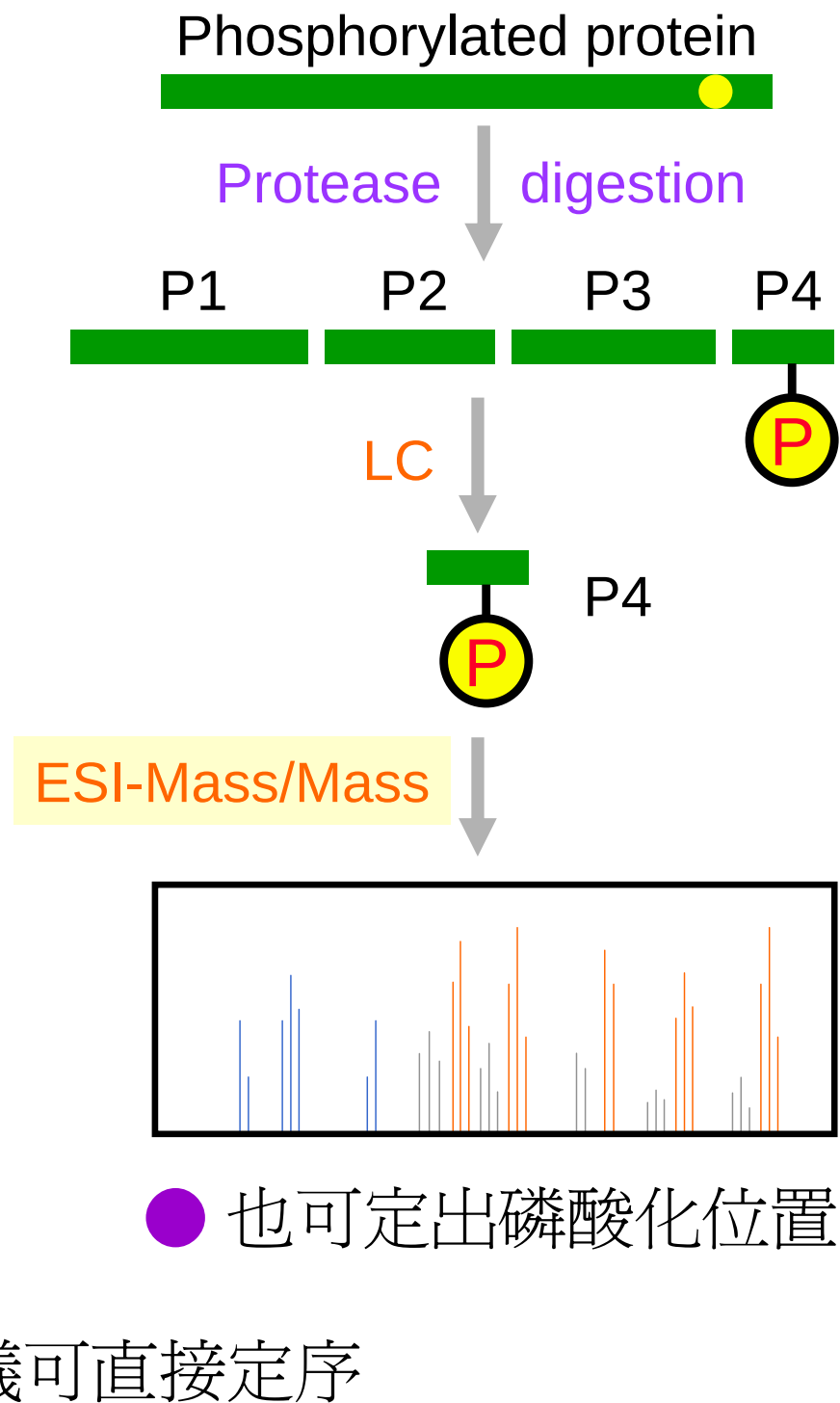
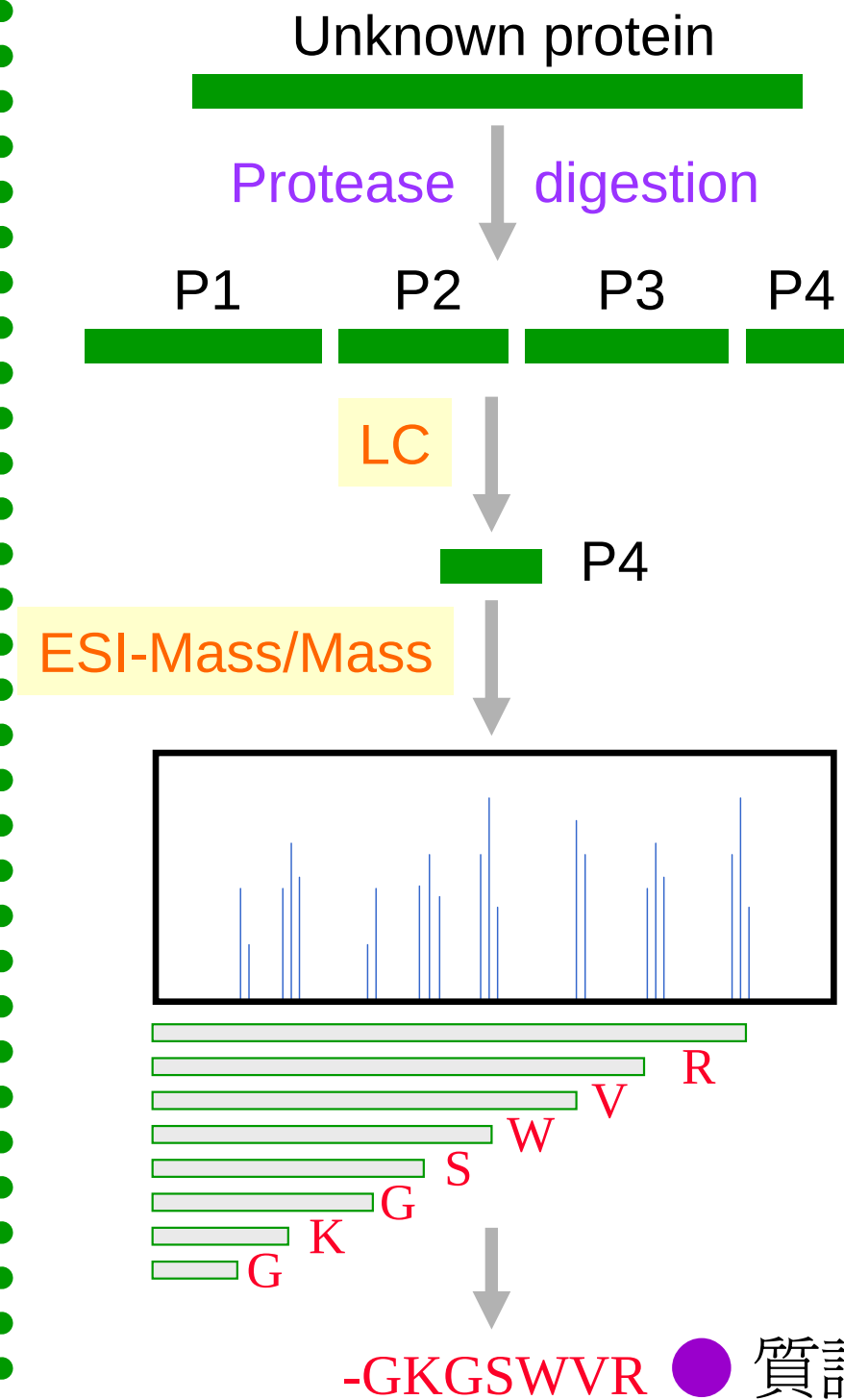


質譜儀可檢定蛋白質身分



● 比對各片段分子量可確定該蛋白質身分

質譜儀可進行蛋白質序列分析



■ 現代蛋白質科技的特點

- 高產能 High-through put
- 快速 High-speed
- 微量 Micro-scaled

